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Chicago, IL
December 6, 2019

“Genetic and Cellular Basis of Lethal
Cardiac Arrhythmia ”

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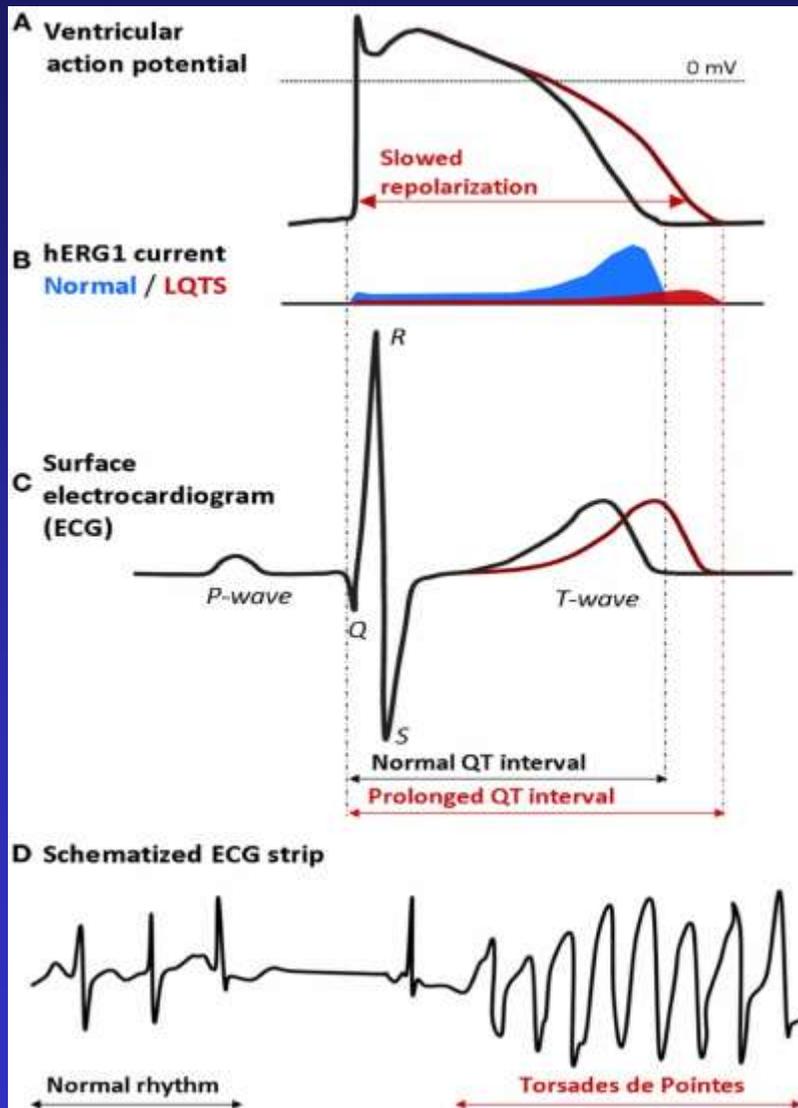


Inherited Cardiac Arrhythmia Syndromes

Common Link: Arrhythmogenic Substrate Develops as a Result of Amplification of Spatial Dispersion of Repolarization

- Long QT Syndrome Preferential prolongation of APD of M cells
- Short QT Syndrome Preferential abbreviation of APD of Epicardium
- Brugada Syndrome Preferential abbreviation of APD of RV epicardium
- Early Repolarization Syndrome Preferential abbreviation of APD in epicardium of the inferior LV

Long QT Syndrome (LQTS)



Diagnosis:

$QTc \geq 450-480$ ms

$QTc > 500$ ms – high risk

Gene Defects Responsible for the Long QT Syndrome

| Chromosome | | Gene | Ion Channel | |
|------------|----|--|--|--------|
| LQT1 | 11 | <i>KCNQ1, KvLQT1</i> | ↓ I _{Ks} | } 90 % |
| LQT2 | 7 | <i>KCNH2, HERG</i> | ↓ I _{Kr} | |
| LQT3 | 3 | <i>SCN5A, Na_v1.5</i> | ↑ Late I _{Na} | |
| LQT4 | 4 | <i>Ankyrin-B, ANK2</i> | ↑ Ca _i , ↑ Late I _{Na} ? | |
| LQT5 | 21 | <i>KCNE1, minK</i> | ↓ I _{Ks} | |
| LQT6 | 21 | <i>KCNE2, MiRP1</i> | ↓ I _{Kr} | |
| LQT7* | 17 | <i>KCNJ2, Kir2.1</i> | ↓ I _{K1} | |
| LQT8** | 6 | <i>CACNA1C, Ca_v1.2</i> | ↑ I _{Ca} | |
| LQT9 | 3 | <i>CAV3, Caveolin-3</i> | ↑ Late I _{Na} | |
| LQT10 | 11 | <i>SCN4B, NavB4</i> | ↑ Late I _{Na} | |
| LQT11 | 7 | <i>AKAP9, Yotiao</i> | ↓ I _{Ks} | |
| LQT12 | 20 | <i>SNTA1, α-1 Syntrophin</i> | ↑ Late I _{Na} | |
| LQT13 | 11 | <i>KCNJ5, Kir3.4</i> | ↓ I _{K-ACh} | |
| LQT14 | 14 | <i>CALM1, Calmodulin</i> | ↑ I _{Ca} , ↑ Late I _{Na} | |
| LQT15 | 2 | <i>CALM2, Calmodulin</i> | ↑ I _{Ca} , ↑ Late I _{Na} | |
| LQT16 | 19 | <i>CALM3, Calmodulin</i> | ↑ I _{Ca} , ↑ Late I _{Na} | |
| LQT17 | 19 | <i>TRPM4</i> , transient receptor potential cation channel | ↓ I _{non-selective cation channel} | |

* Andersen –Tawil Syndrome

** Timothy Syndrome

Congenital Long QT Syndrome (LQTS): Genetics

NIH-funded Clinical Genome Resource (ClinGen) has developed a framework to define and evaluate the clinical validity of gene-disease pairs

Strande et al, Am J Hum Gen 2017

New criteria developed by ACGM (American College of Genetics and Genomics)

Some of the minor genes (**ANK2, KCNE2, SCN4B, AKAP9, SNTA1, and KCNJ5**) have been designated as having **limited- or disputed-evidence** (as monogenic causes).

Minor LQTS genes

| LQTS subtype | Culprit gene |
|--------------|-----------------------|
| LQT1 | KCNQ1 ⁵⁰ |
| LQT2 | KCNH2 ⁵¹ |
| LQT3 | SCN5A ⁵² |
| LQT4 | ANK2 ⁴⁷ |
| LQT5 | KCNE1 ⁵³ |
| LQT6 | KCNE2 ⁵⁴ |
| LQT7 | KCNJ2 ⁵⁵ |
| LQT8 | CACNA1c ⁵⁶ |
| LQT9 | CAV3 ⁴⁶ |
| LQT10 | SCN4B ⁵⁷ |
| LQT11 | AKAP9 ⁴⁸ |
| LQT12 | SNTA1 ³³ |
| LQT13 | KCNJ5 ⁴⁴ |
| LQT14 | CALM1 ²⁷ |
| LQT15 | CALM2 ²⁷ |

Disputed



Strande et al, Am J Hum Gen, 2017

Guidicessi et al, Trend Card Med, 2018

Debate continues as to validity of the ClinGen criteria

It is important to continue to collect both clinical and experimental data concerning their involvement in the pathogenicity of the syndrome which will be reviewed and used to adjust the classifications as necessary

Drugs Associated with LQTS and Torsade de Pointes

Anesthetics

Propofol

Antianginal

Bepidil, Israpidine, Nicardipine

Antiarrhythmic Drugs

Class IA

Quinidine, Procainamide

Disopyramide

Class III

N-acetylprocainamide, sotalol,
Ibutilide, dofetilide

Antibiotics

Erythromycin, Trimethoprim &
Sulfamethazole, Pentamidine,
Clarithromycin

Antihistamines

Terfenadine, Astemizole,
diphenhydramine

Muscle Relaxant

Tizanidine

Antifungal Agents

Ketoconazole

Fluconazole

Itraconazole

Diuretics

Indapamide

Gastrointestinal

Cisapride

Lipid Lowering

Probucol

Psychotropics

Phenothiazines, Tricyclic
antidepressants (Amitriptyline)

Haloperidol, Pimozide

Immunosuppressives

Tacrolimus

Sedative/Hypnotics

Chloral hydrate

Positive Inotropic

DPI 201-106

BDF 9148

Toxins

Anthopleurin-A, ATX-II

Veratridine

Arsenic

Organophosphate
insecticides

Pyrethroids

β – PMTX

Liquid protein diets

Hypokalemia

The genetics underlying acquired long QT syndrome: impact for genetic screening

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Aims

Acquired long QT syndrome (aLQTS) exhibits QT prolongation and Torsades de Pointes ventricular tachycardia triggered by drugs, hypokalaemia, or bradycardia. Sometimes, QTc remains prolonged despite elimination of triggers, suggesting the presence of an underlying genetic substrate. In aLQTS subjects, we assessed the prevalence of mutations in major LQTS genes and their probability of being carriers of a disease-causing genetic variant based on clinical factors.

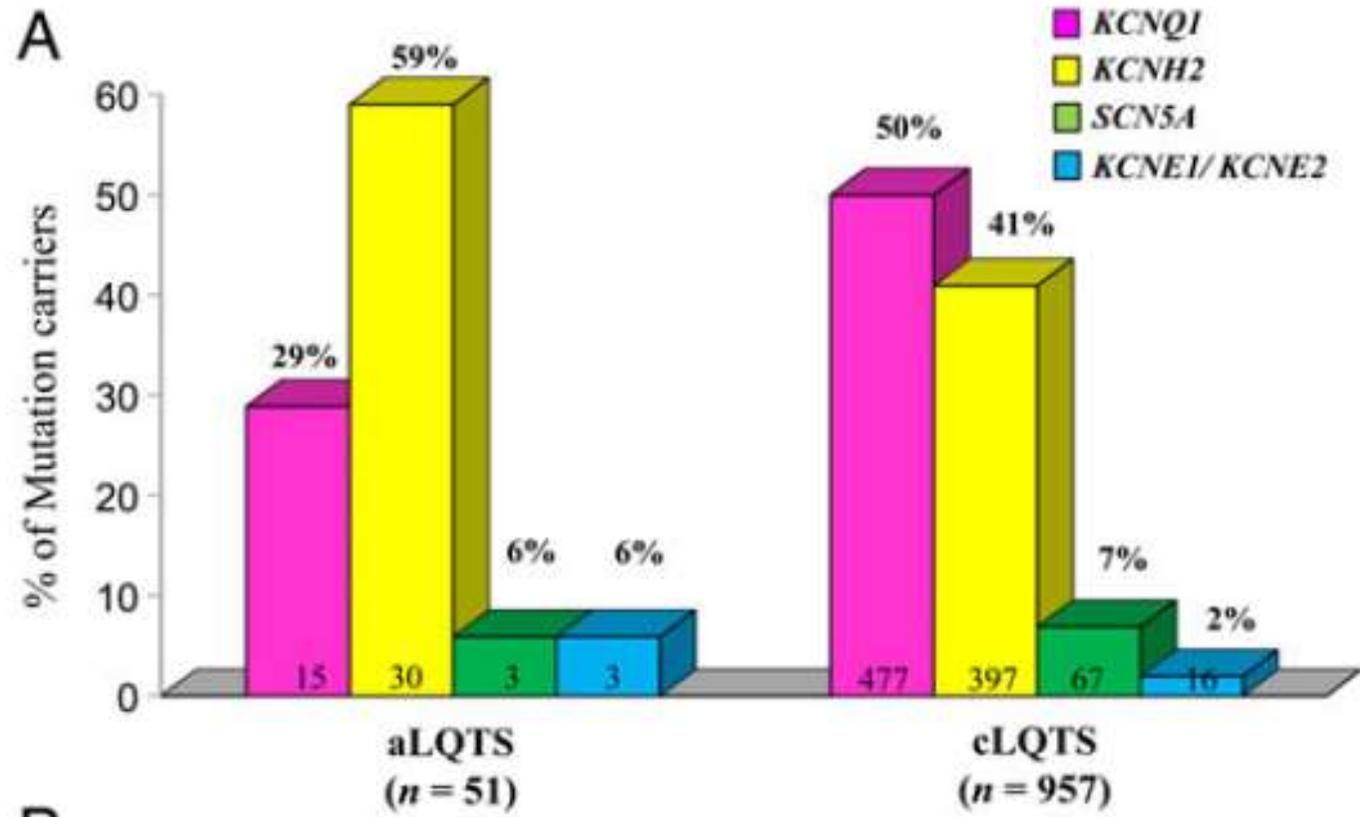
Methods and results

We screened for the five major LQTS genes among 188 aLQTS probands (55 ± 20 years, 140 females) from Japan, France, and Italy. Based on control QTc (without triggers), subjects were designated 'true aLQTS' (QTc within normal limits) or 'unmasked cLQTS' (all others) and compared for QTc and genetics with 2379 members of 1010 genotyped congenital long QT syndrome (cLQTS) families. Cardiac symptoms were present in 86% of aLQTS subjects. Control QTc of aLQTS was 453 ± 39 ms, shorter than in cLQTS (478 ± 46 ms, $P < 0.001$) and longer than in non-carriers (406 ± 26 ms, $P < 0.001$). In 53 (28%) aLQTS subjects, 47 disease-causing mutations were identified. Compared with cLQTS, in 'true aLQTS', *KCNQ1* mutations were much less frequent than *KCNH2* (20% [95% CI 7–41%] vs. 64% [95% CI 43–82%], $P < 0.01$). A clinical score based on control QTc, age, and symptoms allowed identification of patients more likely to carry LQTS mutations.

Conclusion

A third of aLQTS patients carry cLQTS mutations, those on *KCNH2* being more common. The probability of being a carrier of cLQTS disease-causing mutations can be predicted by simple clinical parameters, thus allowing possibly cost-effective genetic testing leading to cascade screening for identification of additional at-risk family members.

Mutations in acquired vs. congenital LQTS

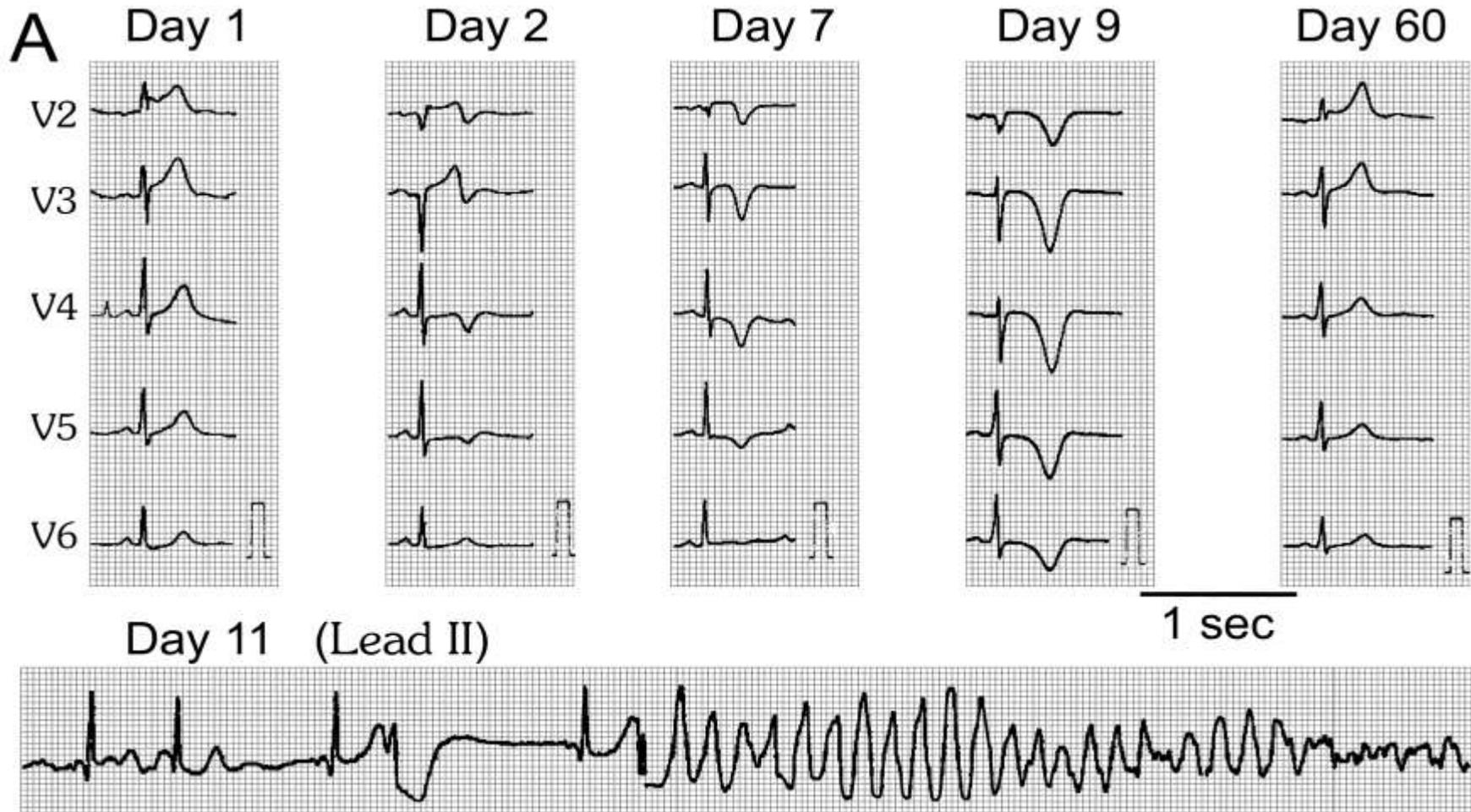


Other Forms of Acquired Long QT Syndrome

- Hypertrophic Cardiomyopathy
- Dilated Cardiomyopathy
- Heart Failure
- Post MI (days 2-11)

- ↓ I_{Kr}
- ↓ I_{Ks}
- ↑ Late I_{Na}
- ↑ I_{Na-Ca}

Post-MI LQTS and TdP



Halkin et al. JACC 38: 1168-74, 2001

Crotti et al. Heart Rhythm, 9:1104-12, 2012

Torsades de pointes following acute myocardial infarction: Evidence for a deadly link with a common genetic variant

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BACKGROUND Although QT prolongation following myocardial infarction (MI) is generally moderate, cases with marked QT prolongation leading to life-threatening torsades de pointes (TdP) have been described.

OBJECTIVE To investigate the genetic substrate of this phenomenon.

METHODS We studied 13 patients who developed TdP in the subacute phase of MI (2–11 days) and a group of 133 ethnically matched controls with uncomplicated MI. Long QT syndrome genes and the *KCNH2*-K897T polymorphism were screened by using denaturing high-performance liquid chromatography plus direct sequencing and a specific TaqMan assay, respectively.

RESULTS Two of the 13 patients (15%) who presented with QT prolongation and TdP were found to carry long QT syndrome mutations (*KCNH2*-R744X and *SCN5A*-E446K). Nine of the remaining 11 patients (82%) carried the *KCNH2*-K897T polymorphism, which was present in 35% of the controls ($P = .0035$). Thus, patients with an acute MI carrying the *KCNH2*-K897T polymorphism had an 8-fold greater risk of experiencing TdP compared with controls (95% confidence interval = 2–40).

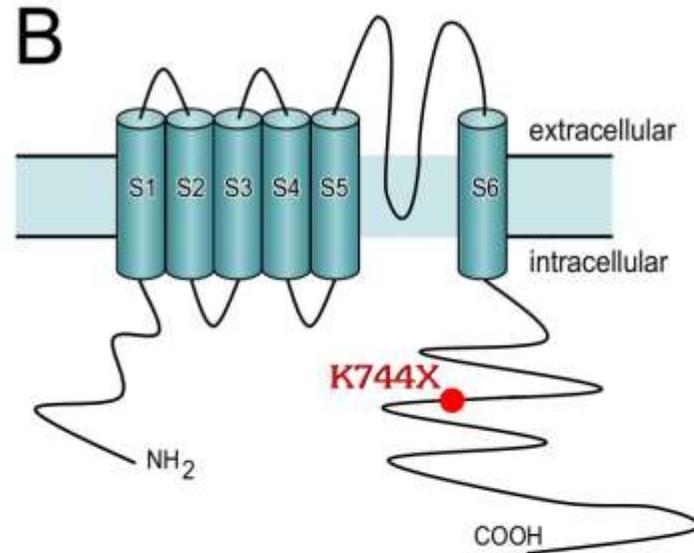
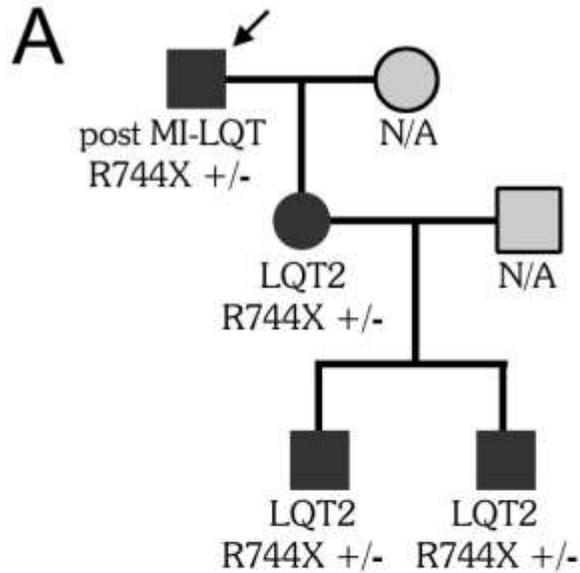
CONCLUSIONS Our data suggest that the common K897T polymorphism is associated with an increased risk of TdP developing in the subacute phase of MI. Our findings support the concept that the electrical remodeling associated with this healing phase of MI may unmask a genetic substrate predisposing to a time-limited development of life-threatening arrhythmias. They also provide the first line of evidence in support of the hypothesis that a common polymorphism, previously described as a modifier of the severity of LQTS, may increase the risk of life-threatening arrhythmias in a much more prevalent cardiac disease such as myocardial infarction.

KEYWORDS Long QT syndrome; Cardiac arrhythmia; Sudden cardiac death; Molecular genetics; Single nucleotide polymorphism

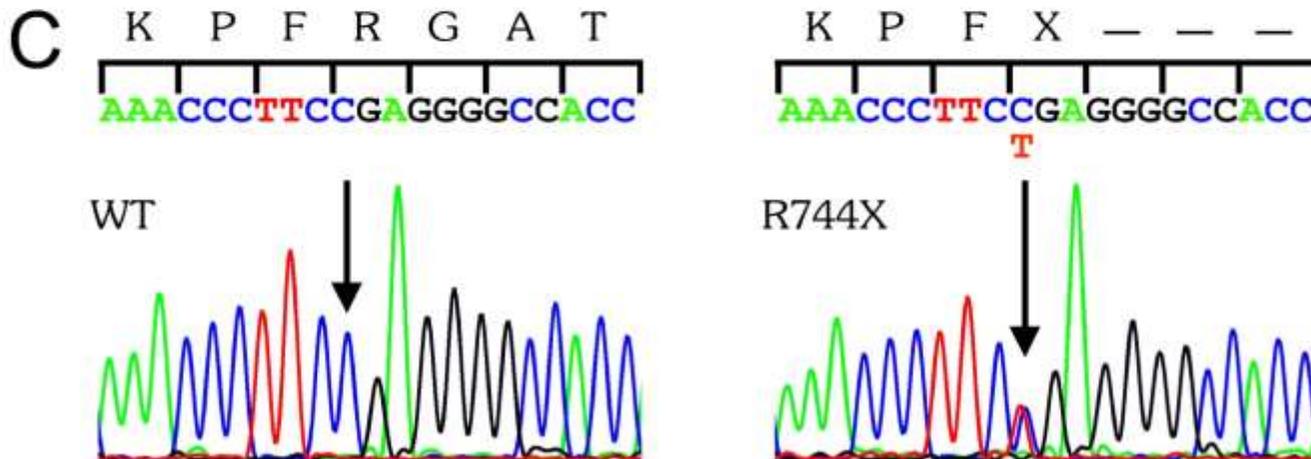
ABBREVIATIONS ECG = electrocardiogram; HERG = human ether-a-go-go-related gene; I_{Kr} = delayed rectifier potassium current; LQTS = long QT syndrome; MI = myocardial infarction; QTc = QT interval corrected for heart rate; SCD = sudden cardiac death; SNP = single nucleotide polymorphism; TdP = torsades de pointes; VF = ventricular fibrillation; WT = wild type

(Heart Rhythm 2012;9:1104–1112) © 2012 Heart Rhythm Society. All rights reserved.

Post-MI LQTS and TdP



KCNH2



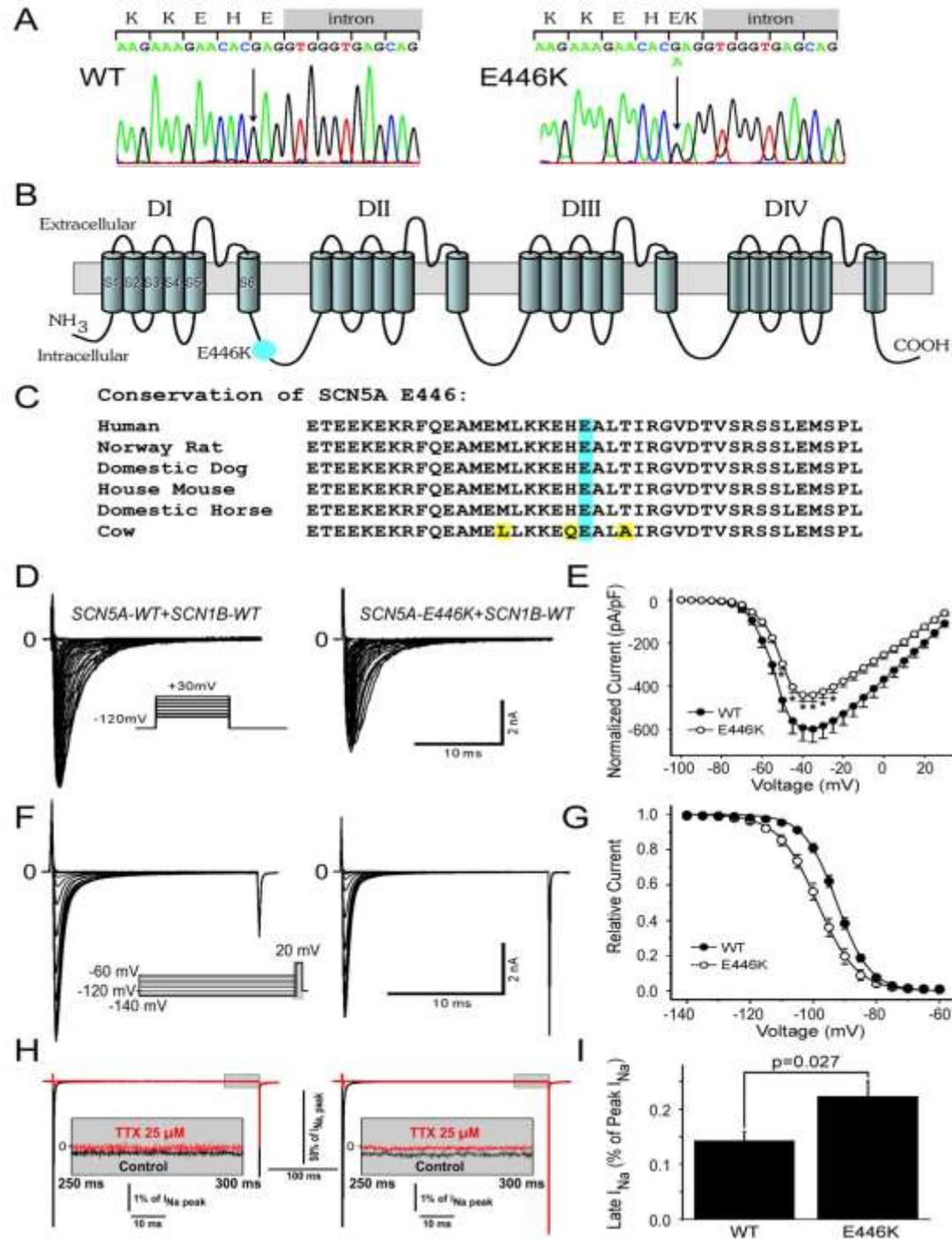
Crotti et al. Heart Rhythm, 9:1104-12, 2012

Post-MI LQTS and TdP

SCN5A-E466K
missense mutation

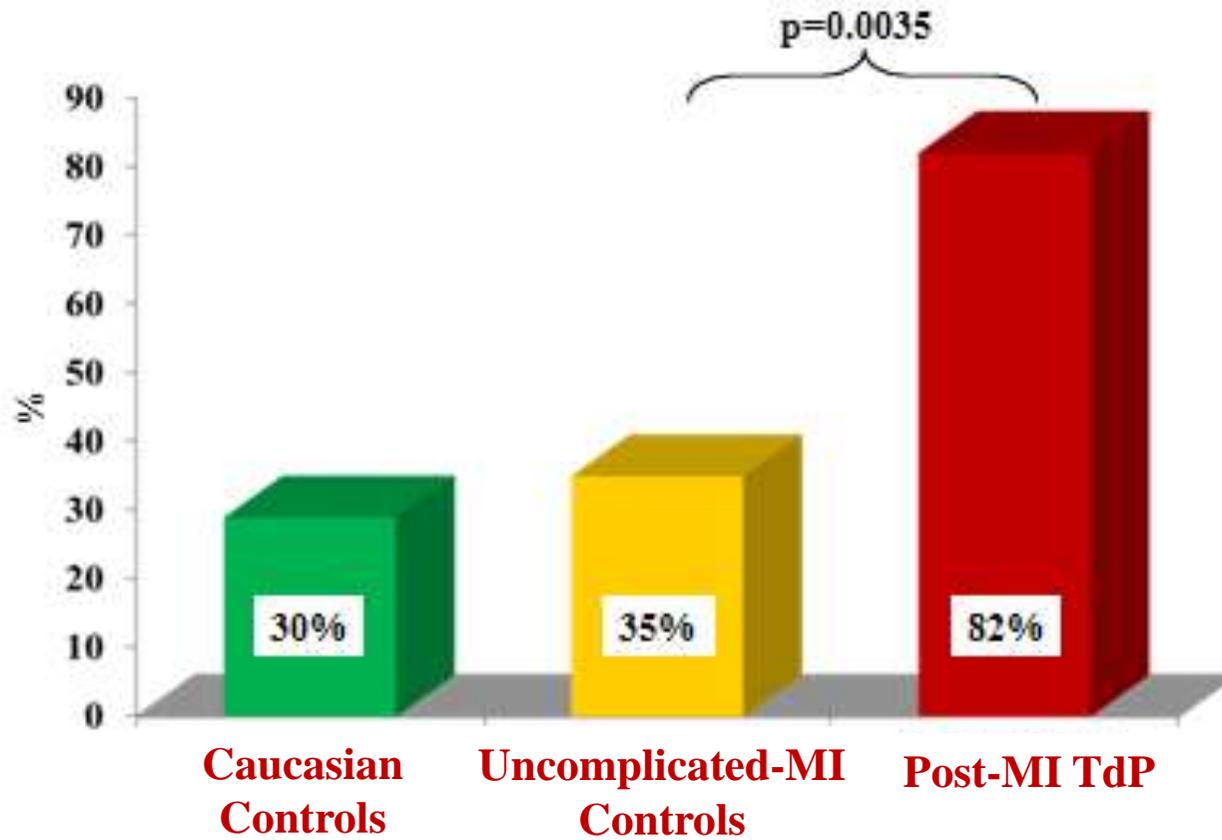
LQT3
Increased I_{Na}

Crotti et al. Heart Rhythm,
9:1104-12, 2012



Post-MI LQTS and TdP

KCNH2-K897T Frequency



Post-MI LQTS and TdP

These data suggest that the common K897T polymorphism is associated with increased risk of TdP developing in the subacute phase of MI.

These findings support the concept that the electrical remodeling associated with this healing phase of MI may unmask a genetic substrate predisposing to a time-limited development of life-threatening arrhythmias.

***KCNH2*-K897T Is a Genetic Modifier of Latent Congenital Long-QT Syndrome**

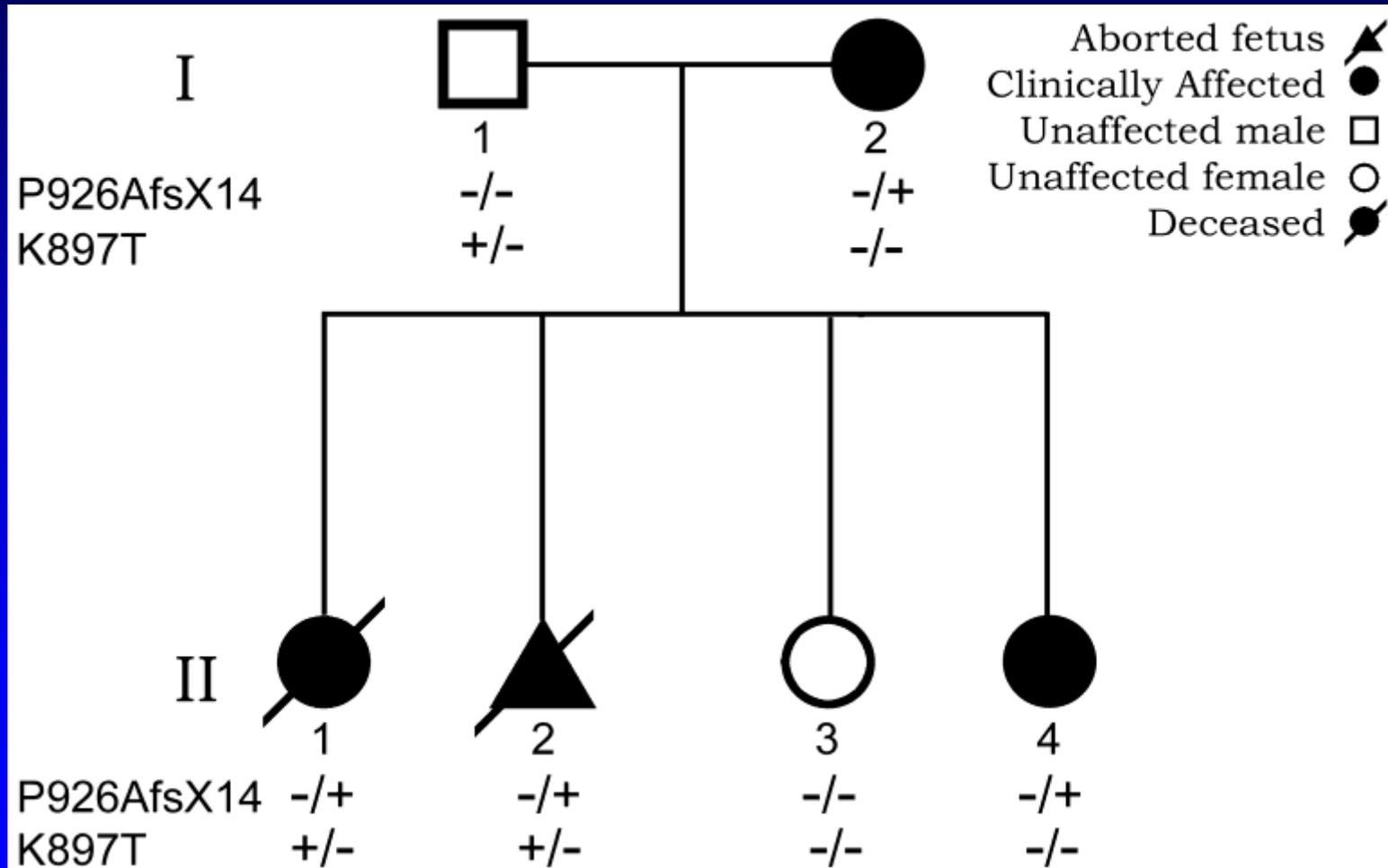
Lia Crotti, MD*; Andrew L. Lundquist, PhD*; Roberto Insolia, BSc; Matteo Pedrazzini, BSc; Chiara Ferrandi, BSc; Gaetano M. De Ferrari, MD; Alessandro Vicentini, MD; Ping Yang, PhD; Dan M. Roden, MD; Alfred L. George, Jr, MD; Peter J. Schwartz, MD

Background—Clinical heterogeneity among patients with long-QT syndrome (LQTS) sharing the same disease-causing mutation is usually attributed to variable penetrance. One potential explanation for this phenomenon is the coexistence of modifier gene alleles, possibly common single nucleotide polymorphisms, altering arrhythmia susceptibility. We demonstrate this concept in a family segregating a novel, low-penetrant *KCNH2* mutation along with a common single nucleotide polymorphism in the same gene.

Methods and Results—The proband is a 44-year-old white woman with palpitations associated with presyncope since age 20, who presented with ventricular fibrillation and cardiac arrest. Intermittent QT prolongation was subsequently observed (max QTc, 530 ms), and LQT2 was diagnosed after the identification of a missense *KCNH2* mutation (A1116V) altering a conserved residue in the distal carboxyl-terminus of the encoded HERG protein. The proband also carried the common *KCNH2* polymorphism K897T on the nonmutant allele. Relatives who carried A1116V without K897T were asymptomatic, but some exhibited transient mild QTc prolongation, suggesting latent disease. Heterologous expression studies performed in cultured mammalian cells and using bicistronic vectors linked to different fluorescent proteins demonstrated that coexpression of A1116V with K897T together resulted in significantly reduced current amplitude as compared with coexpression of either allele with WT-HERG. Thus, the presence of *KCNH2*-K897T is predicted to exaggerate the I_{Kr} reduction caused by the A1116V mutation. These data explain why symptomatic LQTS occurred only in the proband carrying both alleles.

Conclusions—We have provided evidence that a common *KCNH2* polymorphism may modify the clinical expression of a latent LQT2 mutation. A similar mechanism may contribute to the risk for sudden death in more prevalent cardiac diseases. (*Circulation*. 2005;112:1251-1258.)

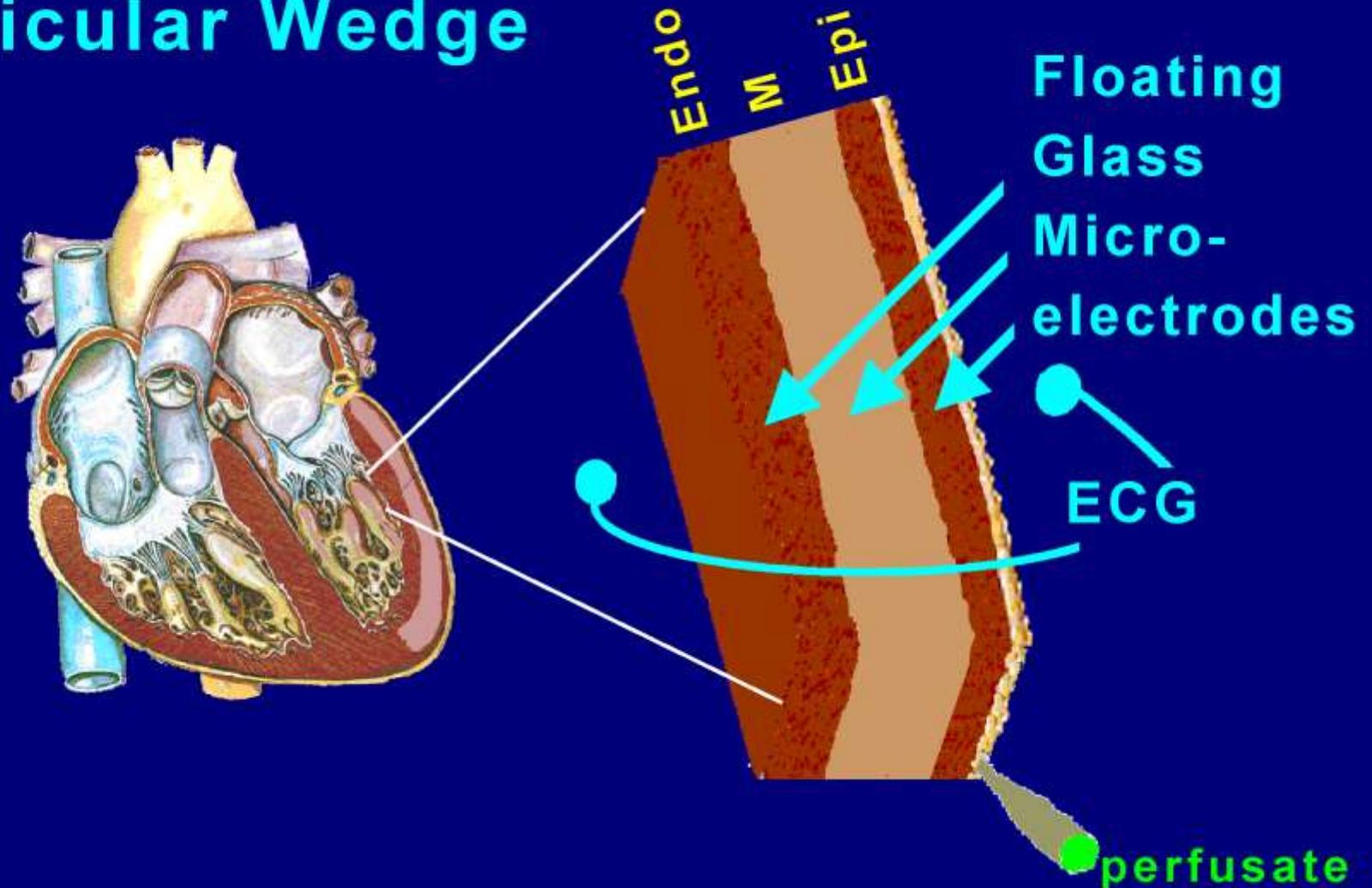
A Common Single Nucleotide Polymorphism (K897T) Can Exacerbate Long QT Type 2 Syndrome Leading to Sudden Infant Death



Nof et al. Circulation Cardiovascular Genetics, 3:199-206, 2010

Arterially Perfused Left Ventricular Wedge

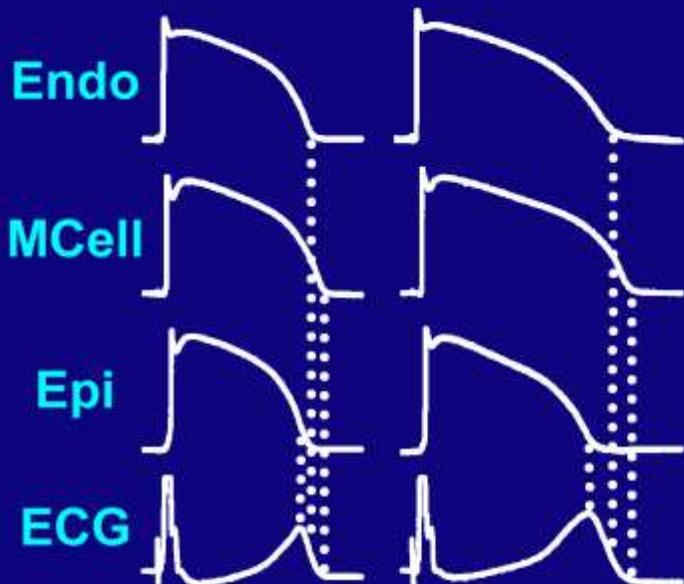
- LQT1
- LQT2
- LQT3
- LQT5
- LQT6
- LQT7
- LQT8



LQT1

Control

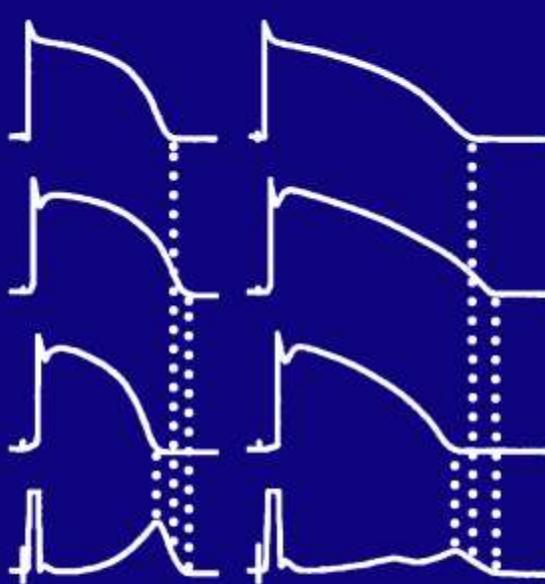
Chromanol 293B (30 μ M)
+ Isoproterenol (100 nM)



LQT2

Control

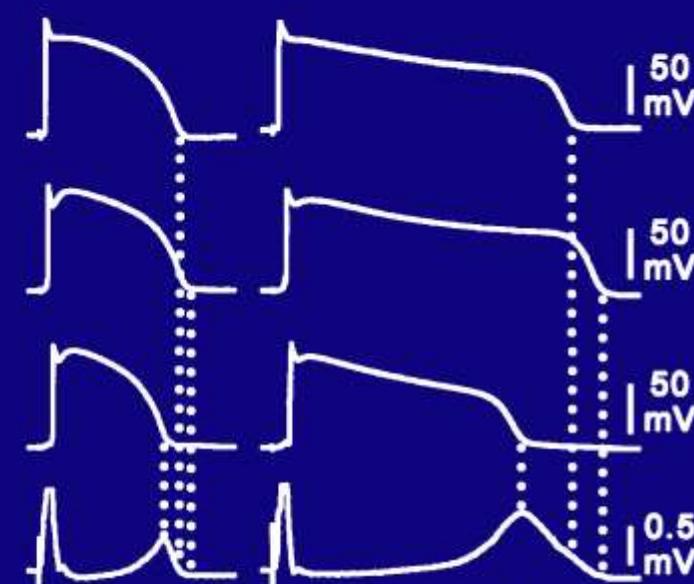
d-Sotalol (100 μ M)
+ low $[K^+]_0$



LQT3

Control

ATX-II (20 nM)

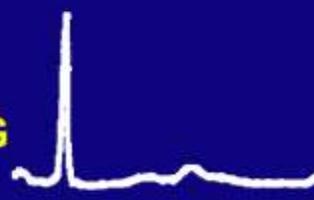


Clinical
ECG
(V5)

KvLQT1



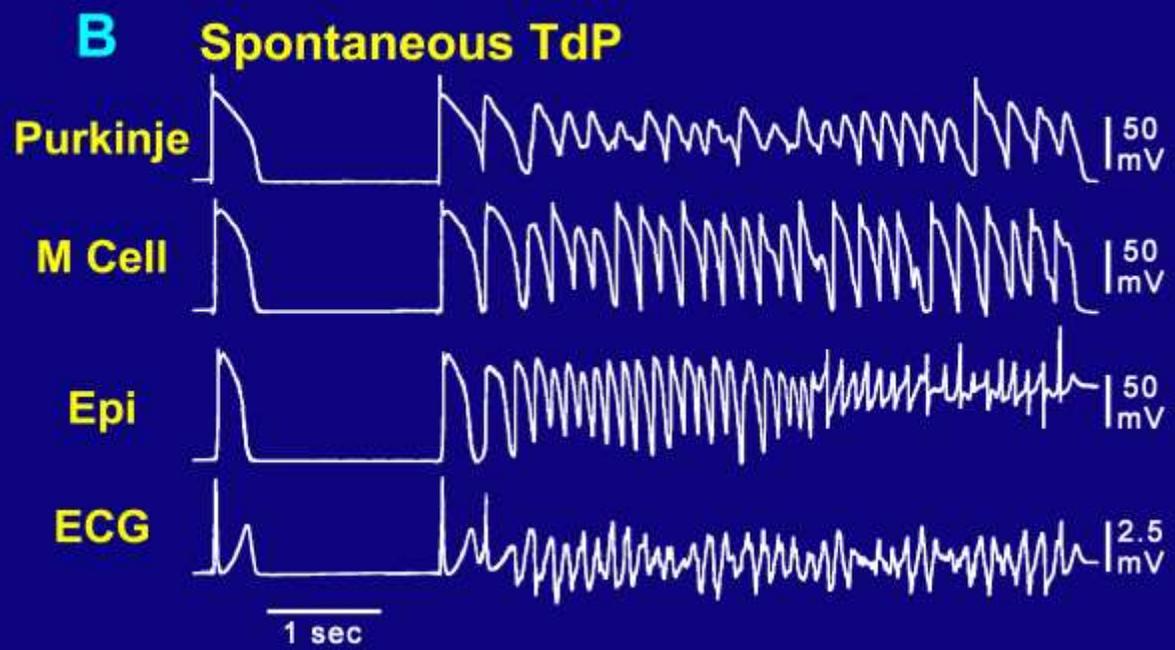
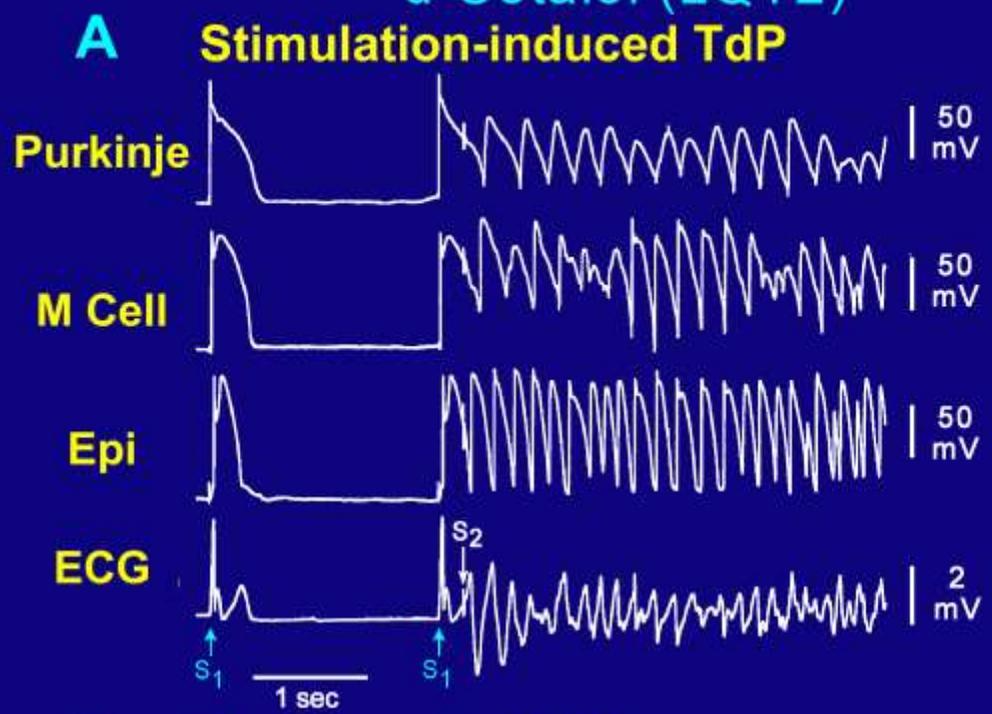
HERG



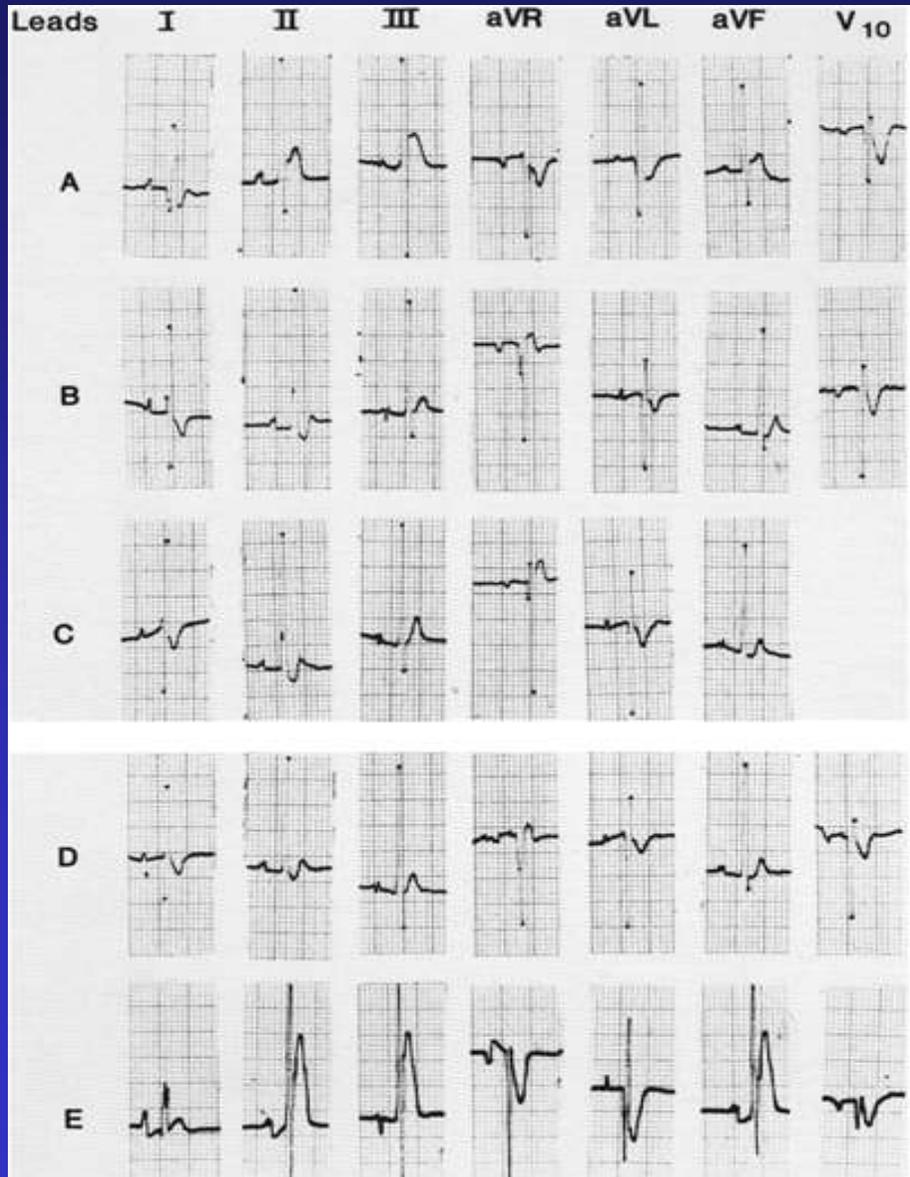
SCN5A



d-Sotalol (LQT2)



Short QT Syndrome

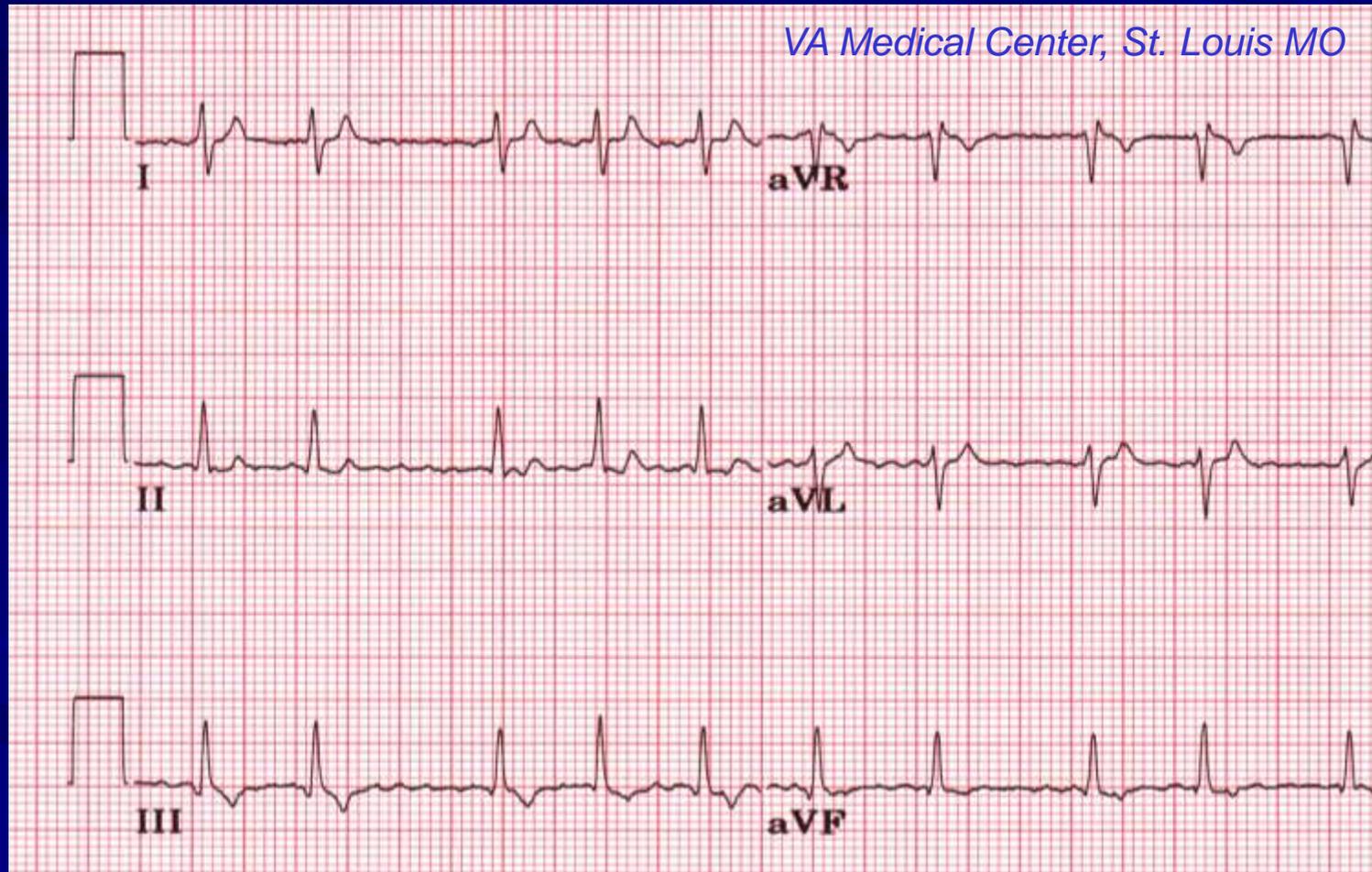


Eastern Grey Kangaroo



Rezakhani A et al. , Austr Vet J 1986

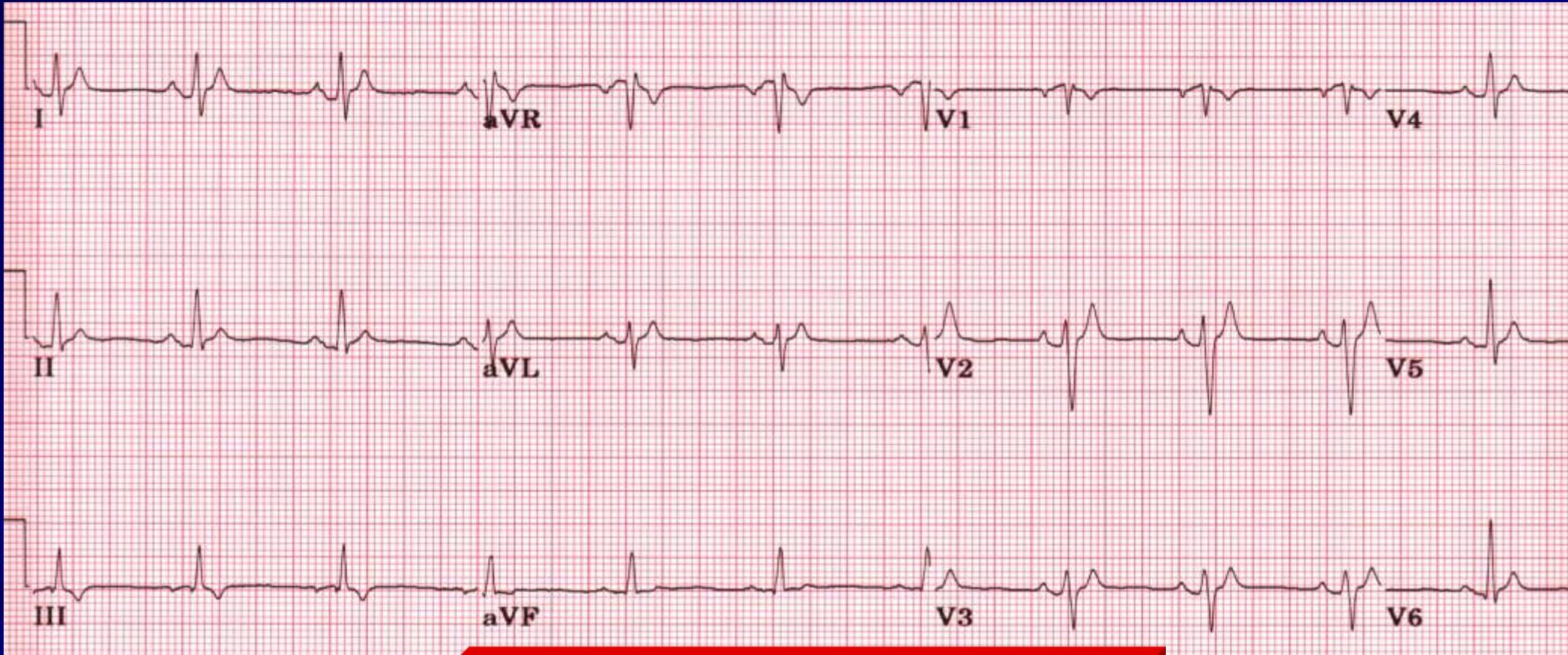
The history of **Short QT Syndrome** started with this ECG of a 17 year old female who presented with Atrial Fibrillation at the Clinic of Preben Bjerregaard in March, 1999



Gussak et al. Cardiology 2000; 94: 99–102

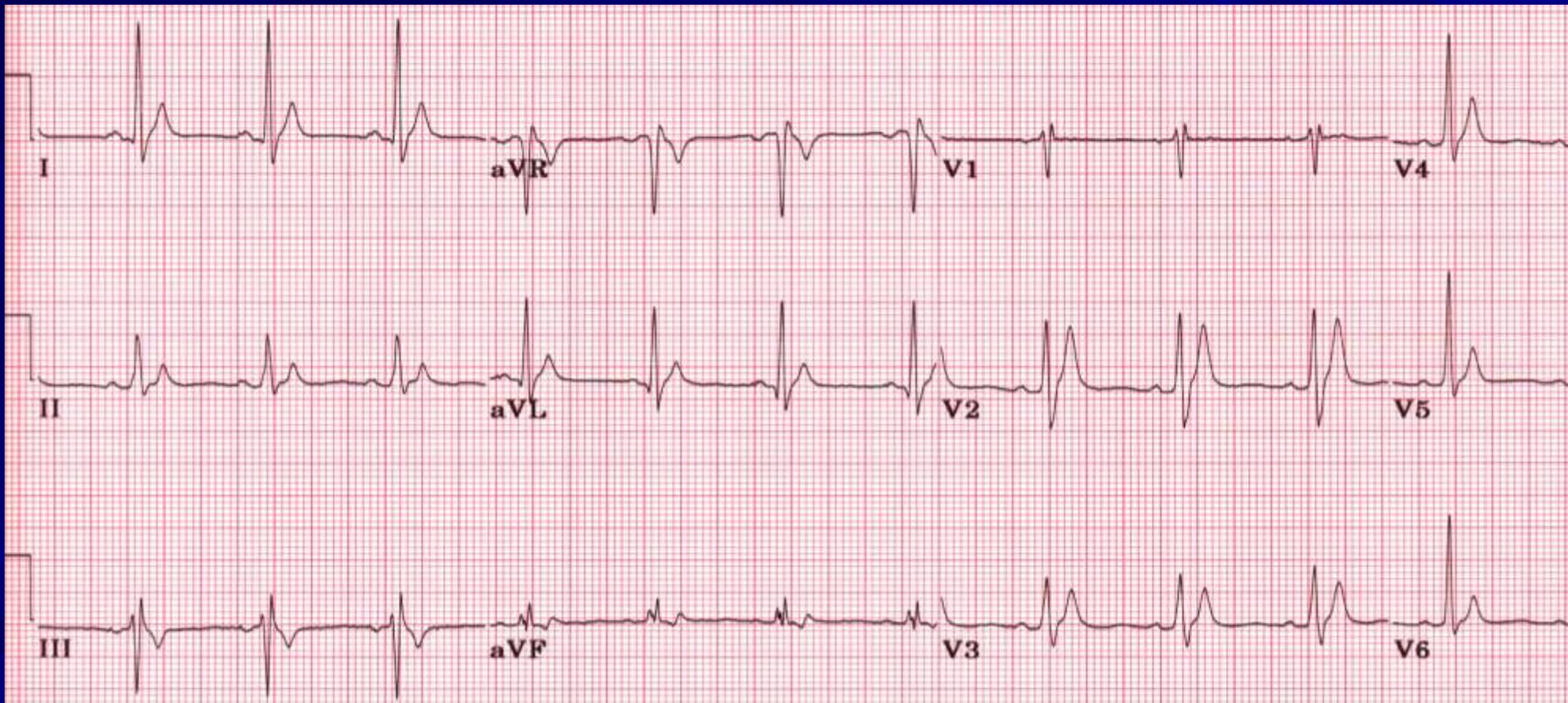
ECG of Index Patient

Sinus Rhythm and Short QT Intervals



QT = 230 msec, QTc = 300 msec

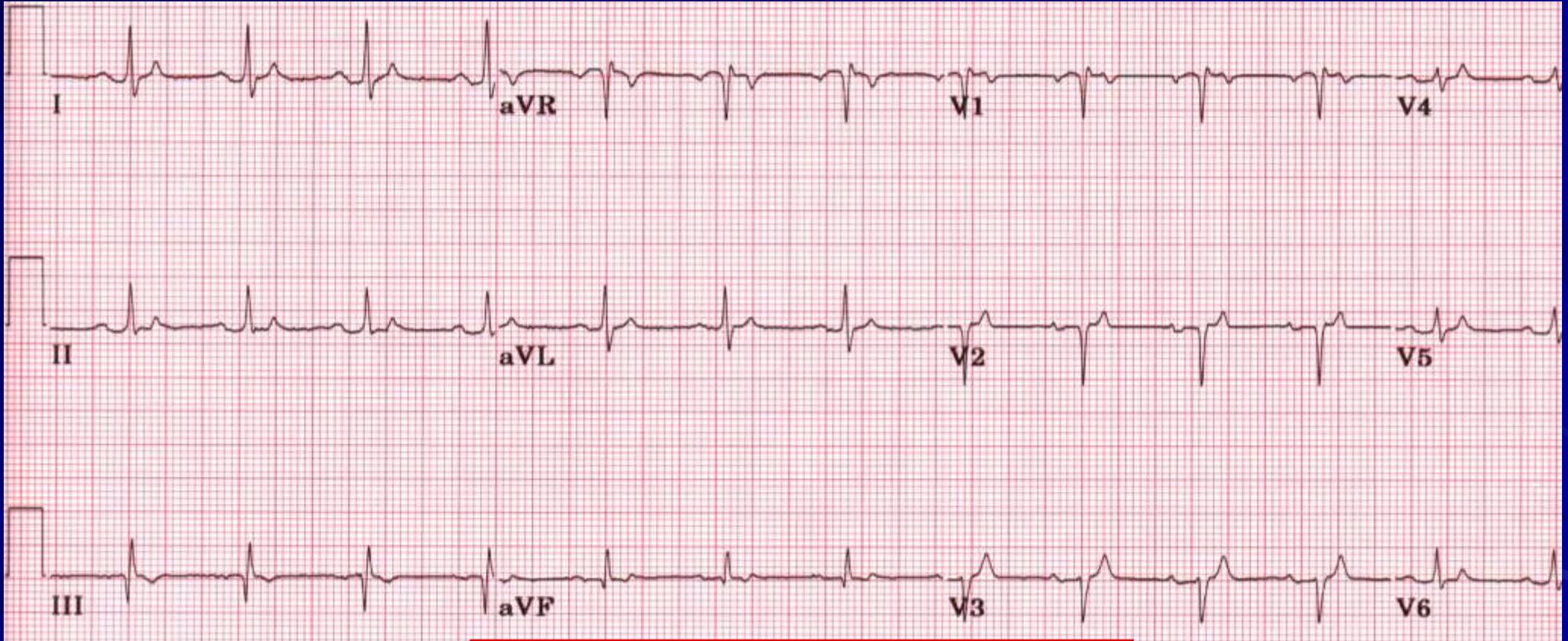
ECG of Brother of Index Patient *Sinus Rhythm and Short QT Intervals*



QT = 245 msec, QTc = 267 msec

ECG of Mother of Index Patient

Sinus Rhythm and Short QT Intervals



QT = 235 msec, QTc 289 msec

Short QT Syndrome

Idiopathic Short QT Interval: A New Clinical Syndrome?

Ihor Gussak^a Pedro Brugada^b Josep Brugada^c R. Scott Wright^a
Stephen L. Kopecky^a Bernard R. Chaitman^d Preben Bjerregaard^d

Cardiology 2000;
94(2):99–102.

ECG Phenomenon of Idiopathic and Paradoxical Short QT Intervals

Cardiac Electrophysiology Review 2002;6:49–53

*Ihor Gussak,¹ Pedro Brugada,² Josep Brugada,³
Charles Antzelevitch,⁴ Mary Osbakken¹
and Preben Bjerregaard⁵*

CER;
2002

Short QT Syndrome A Familial Cause of Sudden Death

Fiorenzo Gaita, MD; Carla Giustetto, MD; Francesca Bianchi, MD; Christian Wolpert, MD;
Rainer Schimpf, MD; Riccardo Riccardi, MD; Stefano Grossi, MD;
Elena Richiardi, MD; Martin Borggreffe, MD

Circulation
2003; 108:
965-70

Diagnosis of Short QT Syndrome

2015 ESC Guidelines for the management of patients with ventricular arrhythmias and the prevention of sudden cardiac death

The Task Force for the Management of Patients with Ventricular Arrhythmias and the Prevention of Sudden Cardiac Death of the European Society of Cardiology (ESC)

Endorsed by: Association for European Paediatric and Congenital Cardiology (AEPC)

Authors/Task Force Members: Silvia G. Priori* (Chairperson) (Italy), Carina Blomström-Lundqvist* (Co-chairperson) (Sweden), Andrea Mazzanti† (Italy), Nico Blom^a (The Netherlands), Martin Borggrefe (Germany), John Camm (UK), Perry Mark Elliott (UK), Donna Fitzsimons (UK), Robert Hatala (Slovakia), Gerhard Hindricks (Germany), Paulus Kirchhof (UK/Germany), Keld Kjeldsen (Denmark), Karl-Heinz Kuck (Germany), Antonio Hernandez-Madrid (Spain), Nikolaos Nikolaou (Greece), Tone M. Norekvål (Norway), Christian Spaulding (France), and Dirk J. Van Veldhuisen (The Netherlands)

| Recommendations | Class ^a | Level ^b | Ref. ^c |
|--|--------------------|--------------------|-----------------------|
| SQTS is diagnosed in the presence of a QTc ≤ 340 ms. | I | C | This panel of experts |
| SQTS should be considered in the presence of a QTc ≤ 360 ms and one or more of the following: (a) A confirmed pathogenic mutation (b) A family history of SQTS (c) A family history of sudden death at age < 40 years (d) Survival from a VT/VF episode in the absence of heart disease. | IIa | C | This panel of experts |

QTc = corrected QT; SQTS = short QT syndrome; VF = ventricular fibrillation; VT = ventricular tachycardia.

^aClass of recommendation.

^bLevel of evidence.

^cReference(s) supporting recommendations.



Tall peaked T waves
Positive or Negative

↑ T_{peak-Tend} Interval

= ↑ Transmural
Dispersion of
Repolarization (TDR)

Short QT Syndrome

| | QTc (ms) | Gene (Cardiac Ion Channel) | Reference |
|--------------|-------------|---|--|
| SQT 1 | 286 ± 6 | KCNH2 (I _{Kr}) ↑ | Brugada et al., Circulation 109:30, 2004 |
| SQT 2 | 302 | KCNQ1 (I _{Ks}) ↑ | Belloq et al., Circulation 109:2394, 2004, |
| SQT 3 | 315 - 330 | KCNJ2 (I _{K1}) ↑ | Priori et al., Circulation Research 96: 800, 2005 |
| SQT 4 | 331 - 370 | CACNB2b (I _{Ca}) ↓ | Antzelevitch et al. Circulation 115:442, 2007 |
| SQT 5 | 346-360 | CACNA1C (I _{Ca}) ↓ | Antzelevitch et al. Circulation 115: 442, 2007 |
| SQT 6 | 330 | CACNA2D1 (I _{Ca}) ↓ | Templin et al., European Heart Journal, 32:1077-88, 2011 |
| SQT 7 | 282 - 340 | SLC22A5 (↓carnitine - ↑ I _{Kr}) | Roussel et al., Heart Rhythm 13:165-174, 2016 |
| SQT8 | 340 | SLC4A3 (↑pHi - ↓Cl _i) | Thorsen et al., Nature Comm. 8:1696, 2017 |

Calcium channel mutations often produce a combined SQTs/BrS phenotype

Carnitine deficiency induces a short QT syndrome



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From the ^{}INSERM U1046, CNRS UMR 9214, Université de Montpellier, Montpellier, France, [†]Médecine Pédiatrique, INSERM U1069, CHRU de Tours, Université François Rabelais, Tours, France, [‡]Department of Biochemistry, Faculty of Medicine, Kagoshima University, Kagoshima, Japan, [§]Laboratoire de Biochimie Hormonologie, Hôpital R Debré, AP-HP, Paris, France, and ^{||}Service de Cardiologie, CHRU de Tours, Université François Rabelais, Tours, France.*

BACKGROUND Short QT syndrome is associated with an increased risk of cardiac arrhythmias and unexpected sudden death. Until now, only mutations in genes encoding the cardiac potassium and calcium channels have been implicated in early T-wave repolarization.

OBJECTIVE The purpose of this study was to confirm a relationship between a short QT syndrome and carnitine deficiency.

METHODS We report 3 patients affected by primary systemic carnitine deficiency and an associated short QT syndrome. Ventricular fibrillation during early adulthood was the initial symptom in 1 case. To confirm the relationship between carnitine, short QT syndrome, and arrhythmias, we used a mouse model of carnitine deficiency induced by long-term subcutaneous perfusion of MET88.

RESULTS MET88-treated mice developed cardiac hypertrophy associated with a remodeling of the mitochondrial network. The continuous monitoring of electrocardiograms confirmed a shortening of the QT interval, which was negatively correlated with

the plasma carnitine concentration. As in humans, such alterations coincided with the genesis of ventricular premature beats and ventricular tachycardia and fibrillation.

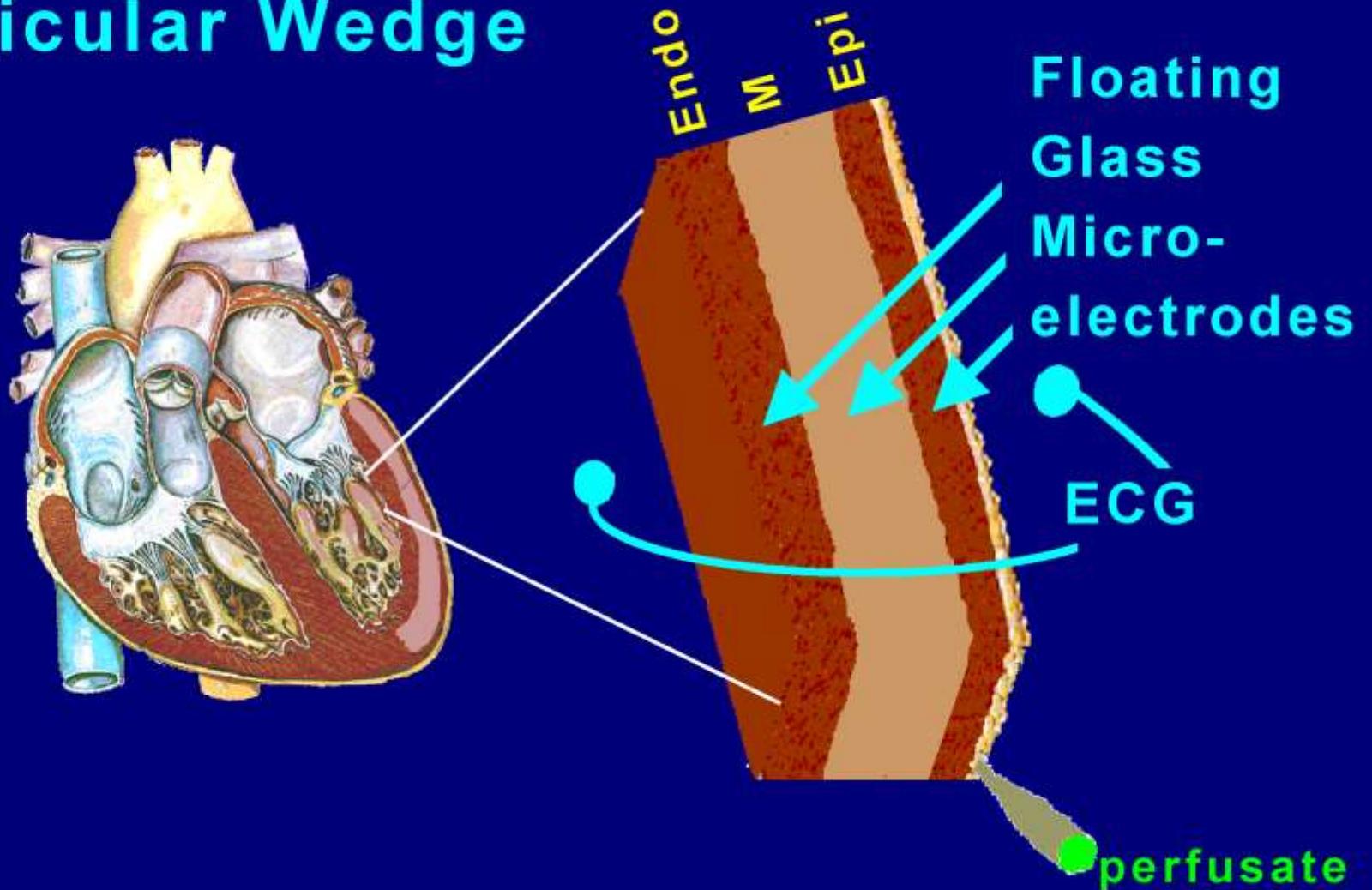
CONCLUSION Altogether, these results suggest that long-chain fatty acid metabolism influence the morphology and the electrical function of the heart.

KEYWORDS Carnitine deficiency; Short QT syndrome; Electrophysiological remodeling; Sudden death; Ventricular arrhythmias

ABBREVIATIONS ECG = electrocardiogram/electrocardiographic; I_{Kr} = rapid potassium current; LCFA = long-chain fatty acid; LV = left ventricular; OCTN2 = organic cation transport Na⁺; PCD = primary carnitine deficiency

(Heart Rhythm 2016;13:165–174) © 2016 Heart Rhythm Society. All rights reserved.

Arterially Perfused Left Ventricular Wedge

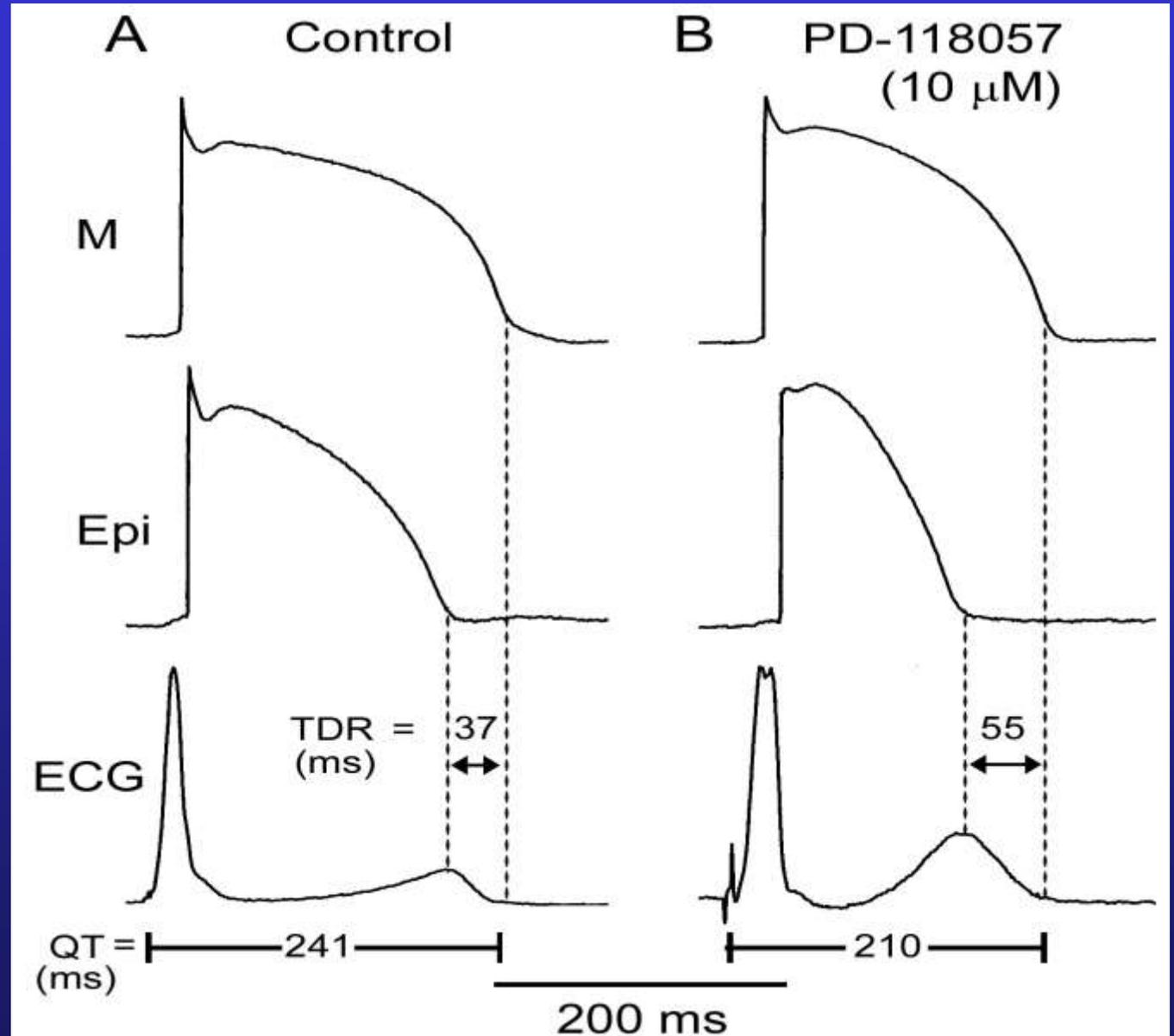
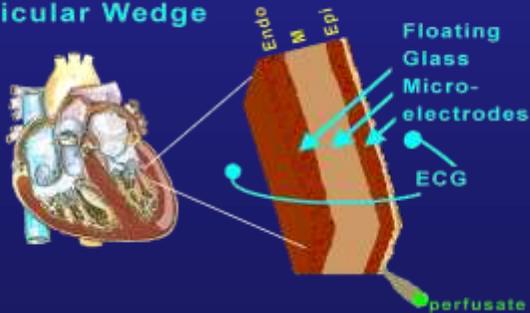


Short QT Syndrome

SQT1

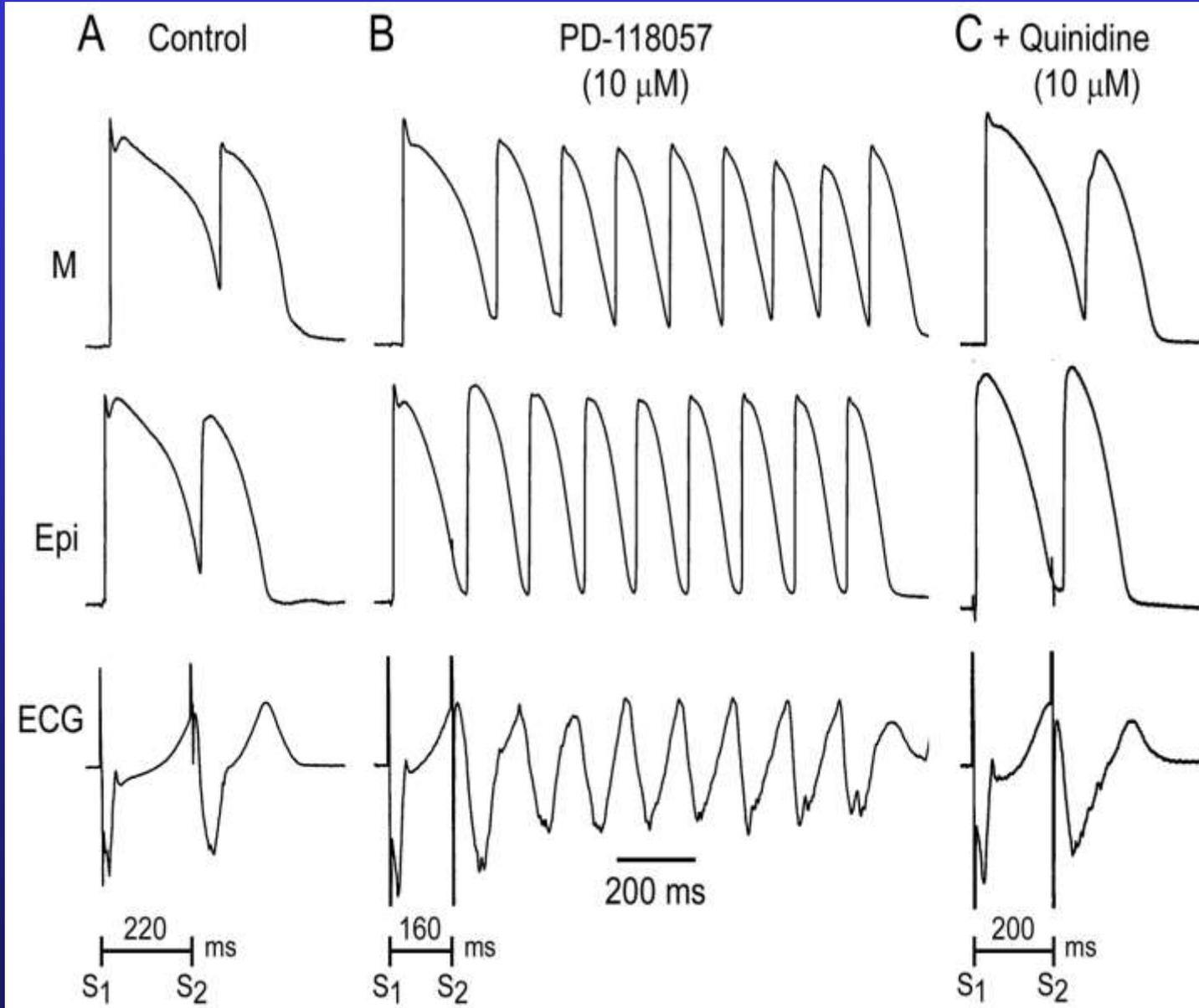
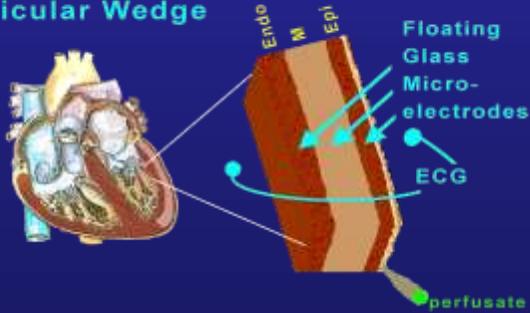
I_{Kr} Agonist

Arterially Perfused Left Ventricular Wedge



Short QT Syndrome

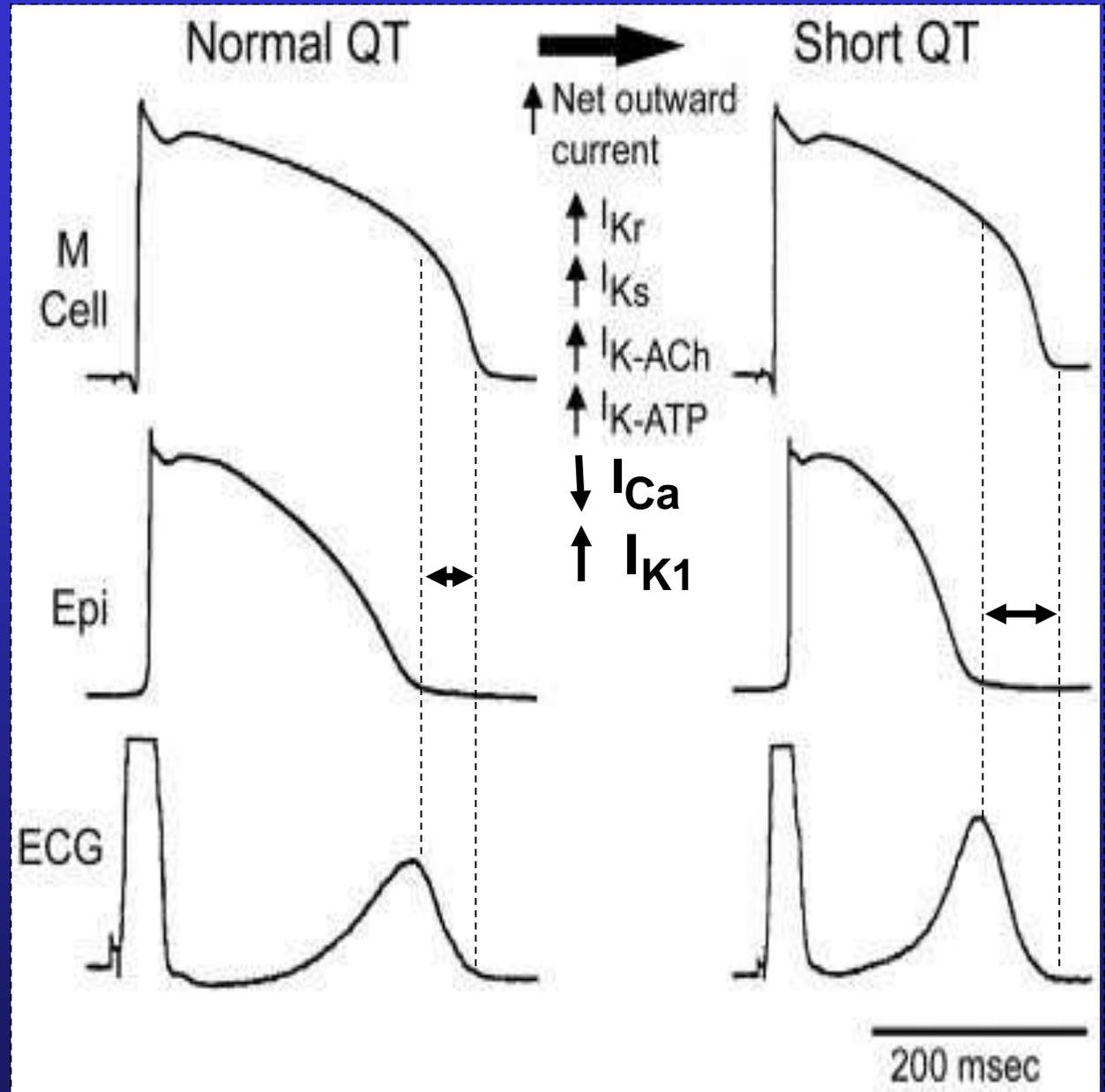
Arterially Perfused Left Ventricular Wedge



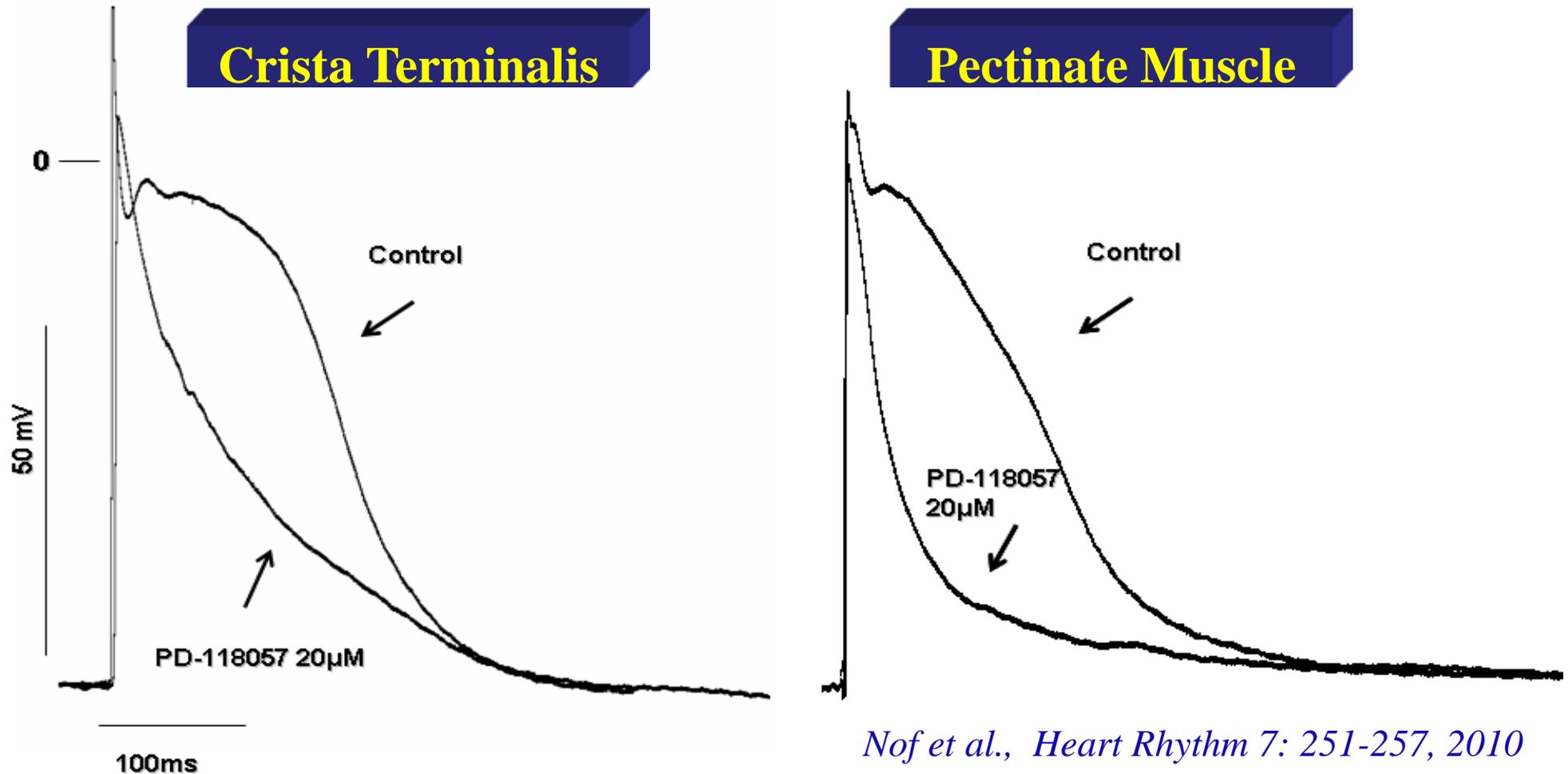
Short QT Syndrome

Arrhythmogenic Mechanism

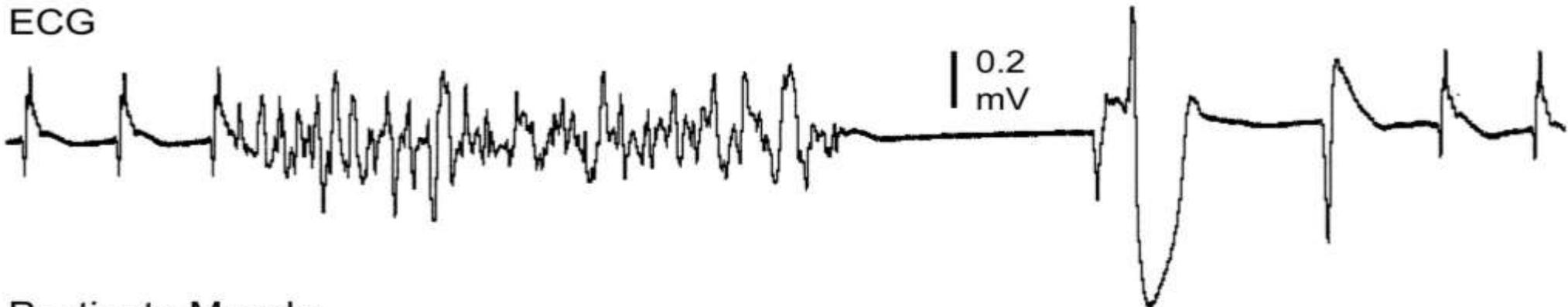
↓ ERP ↑ TDR



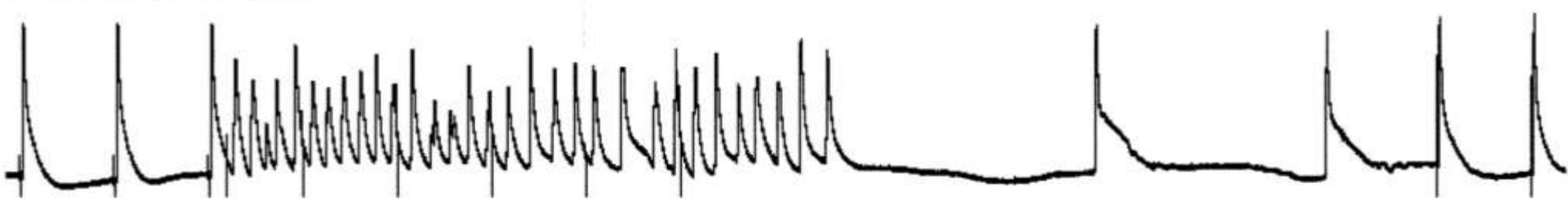
Heterogeneous Abbreviation of APD by I_{Kr} Agonist PD-118057 in Coronary-perfused Canine Right Atrium



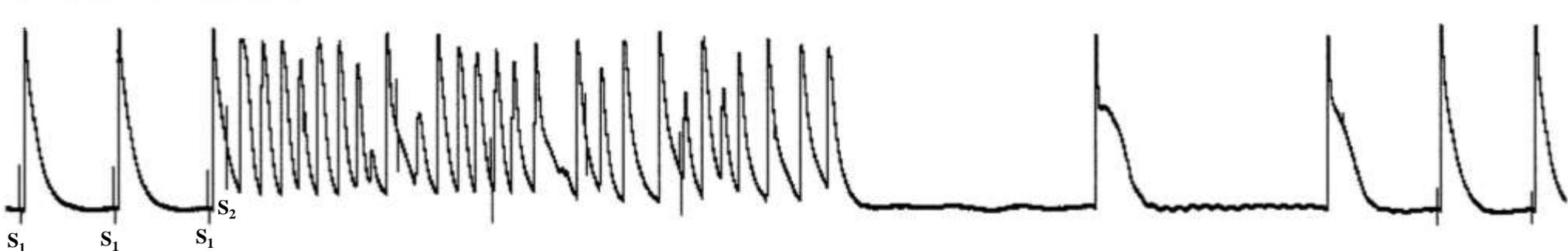
ECG



Pectinate Muscle



Crista Terminalis



S₁ S₁ S₁ S₂

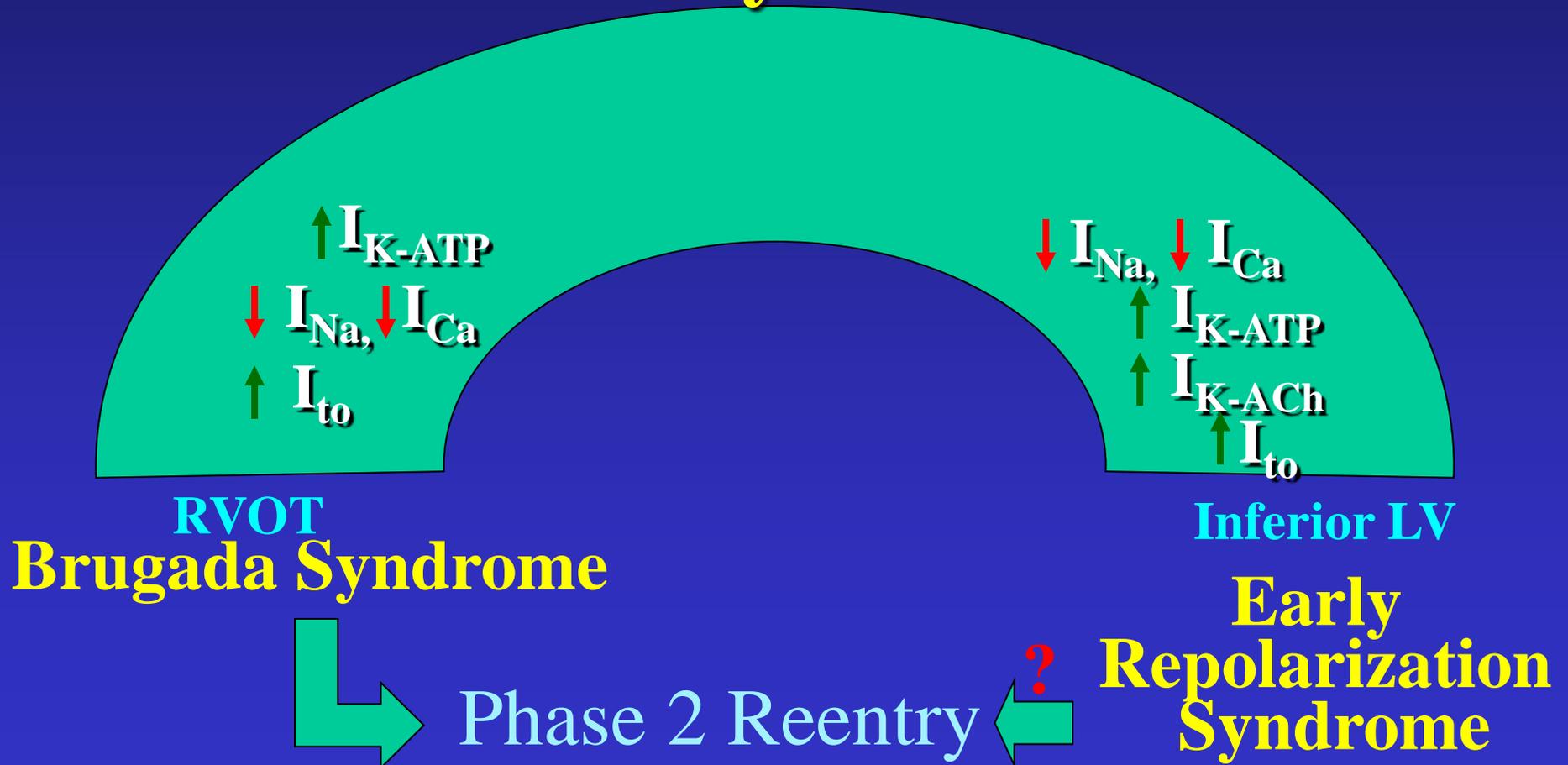


1 sec

Outward shift of repolarizing current during early phase of the action potential



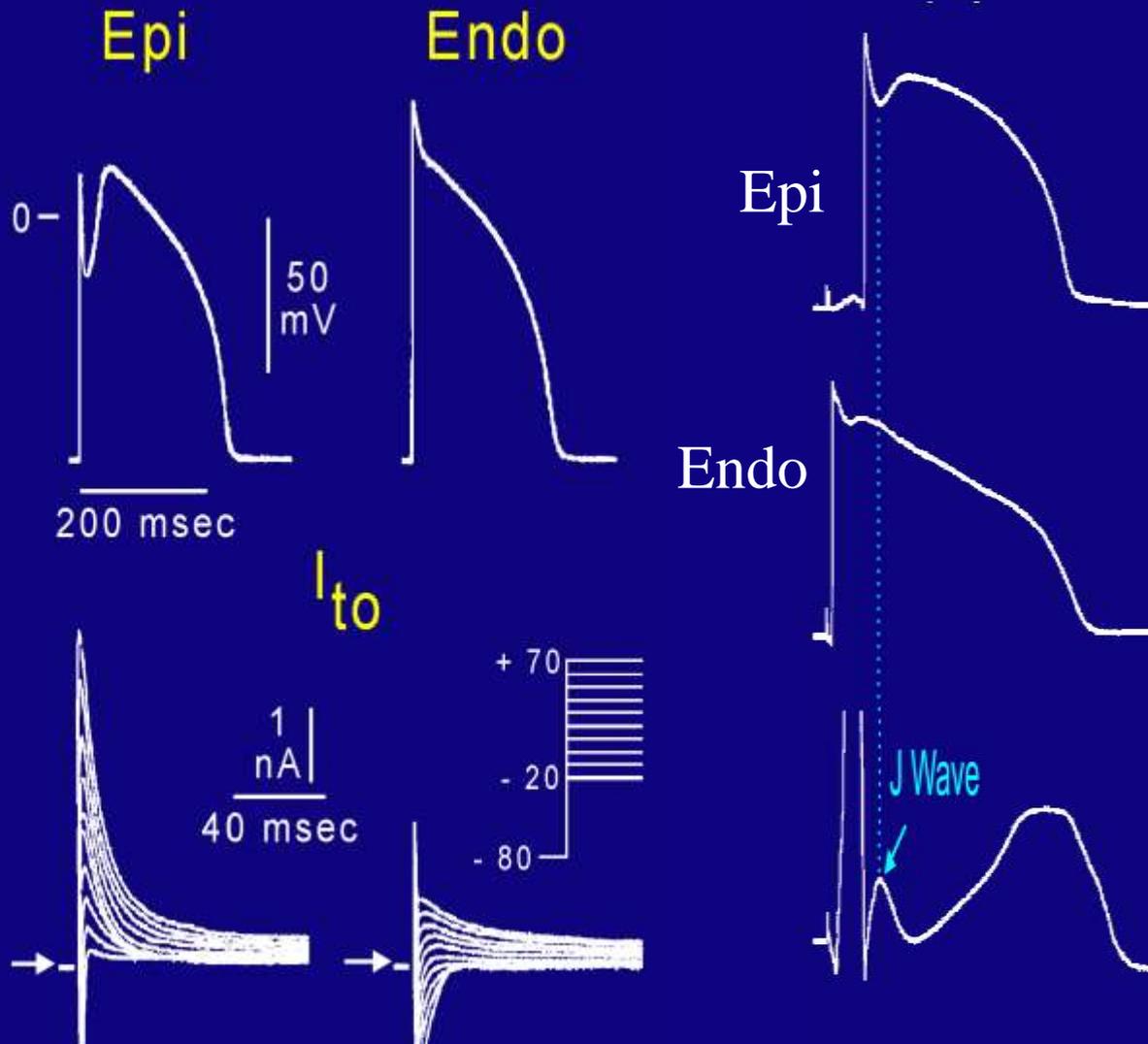
J Wave Syndromes



Continuous Spectrum Between BrS and ERS

- Brugada (BrS) and Early Repolarization (ERS) Syndromes share similar ECG characteristics, clinical outcomes, risk factors and arrhythmic characteristics.
- Although BrS and ERS differ with respect to the magnitude and lead location of abnormal J wave manifestation, they can be considered to represent a continuous spectrum of phenotypic expression, termed J wave syndromes, and to share a common arrhythmic platform related to amplification of I_{to} -mediated J waves.

Cellular Basis for the J Wave



Transmural distribution of the I_{to} -mediated action potential notch is responsible for the inscription of the electrocardiographic J wave

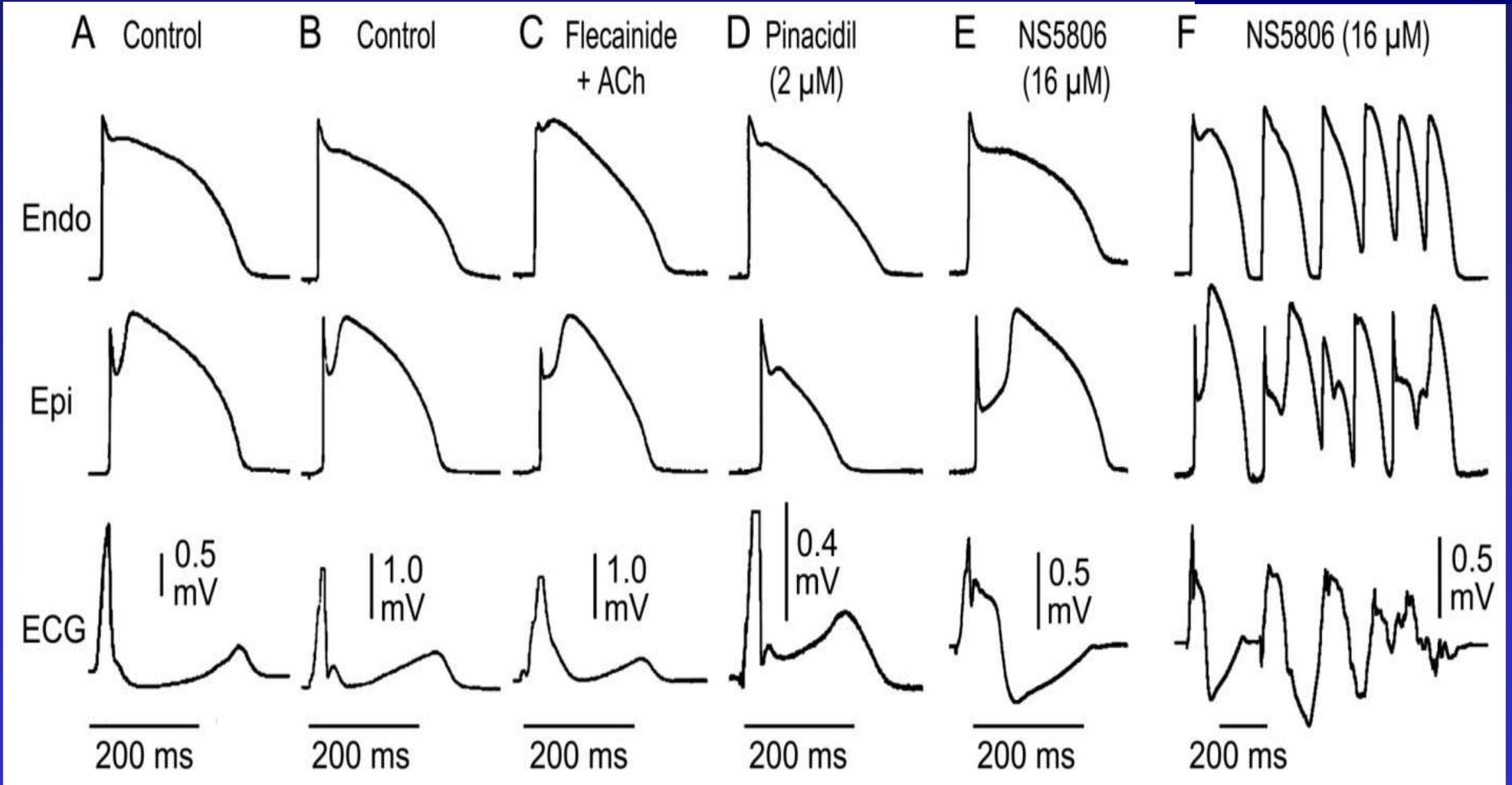
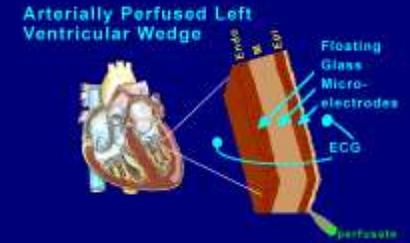
Similarities between Brugada and Early Repolarization Syndromes and Possible Underlying Mechanisms

| | BrS | ERS | Possible Mechanism(s) |
|--|-------------------|-------------------|--|
| Male Predominance | Yes (>75%) | Yes (>80%) | Testosterone modulation of ion currents underlying the epicardial AP notch |
| Average age of first event | 30-50 | 30-50 | |
| Associated with mutations or rare variants in KCNJ8, CACNA1C, CACNB2, CACNA2D, SCN5A, ABCC9, SCN10A | Yes | Yes | Gain of function in outward currents (I_{K-ATP}) or loss of function in inward currents (I_{Ca} or I_{Na}) |
| Dynamicity of ECG | High | High | Autonomic modulation of ion channel currents underlying early phases of the epicardial AP |
| VF often occurs during sleep or at a low level of physical activity | Yes | Yes | Higher level of vagal tone and higher levels of I_{to} at the slower heart rates. |
| VT/VF trigger | Short-coupled PVC | Short-coupled PVC | Phase 2 reentry |
| Ameliorative response to quinidine and bepridil | Yes | Yes | Inhibition of I_{to} and possible vagolytic effect |
| Ameliorative response to Isoproterenol, denopamine and milrinone | Yes | Yes | Increased I_{Ca} and faster heart rate |
| Ameliorative response to cilostazol | Yes | Yes | Increased I_{Ca} , reduced I_{to} and faster heart rate |
| Ameliorative response to pacing | Yes | Yes | Reduced availability of I_{to} due to slow recovery from inactivation |
| Vagally-mediated accentuation of ECG pattern | Yes | Yes | Direct effect to inhibit I_{Ca} and indirect effect to increase I_{to} (due to slowing of heart rate) |
| Effect of sodium channel blockers on unipolar epicardial electrogram | Augmented J waves | Augmented J wave | Outward shift of balance of current in the early phases of the epicardial action potential |
| Fever | Augmented J waves | Augmented J waves | Accelerated inactivation of I_{Na} and accelerated recovery of I_{to} from inactivation. |
| Hypothermia | Augmented J waves | Augmented J waves | Slowed activation of I_{Ca} , leaving I_{to} unopposed. Increased phase 2 reentry, but reduced pVT due to prolongation of APD (Morita et al, 2007) |

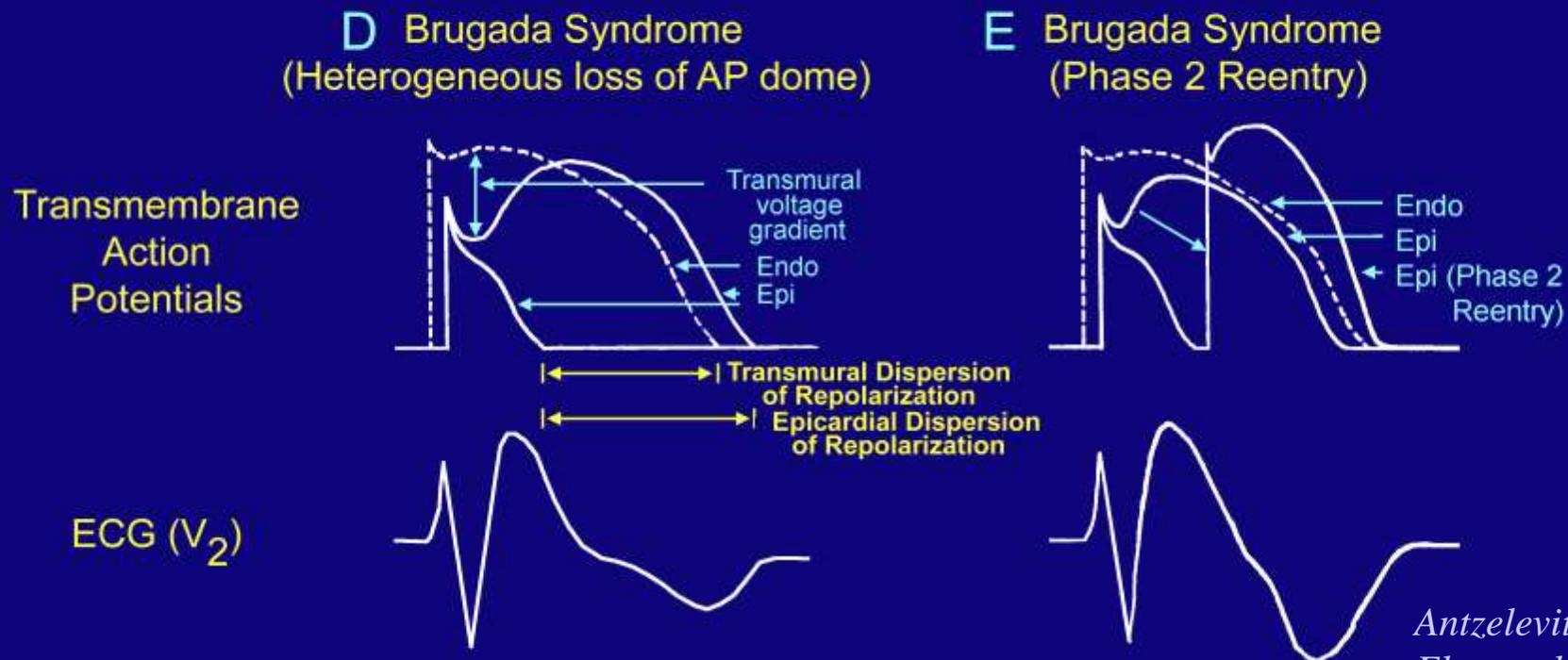
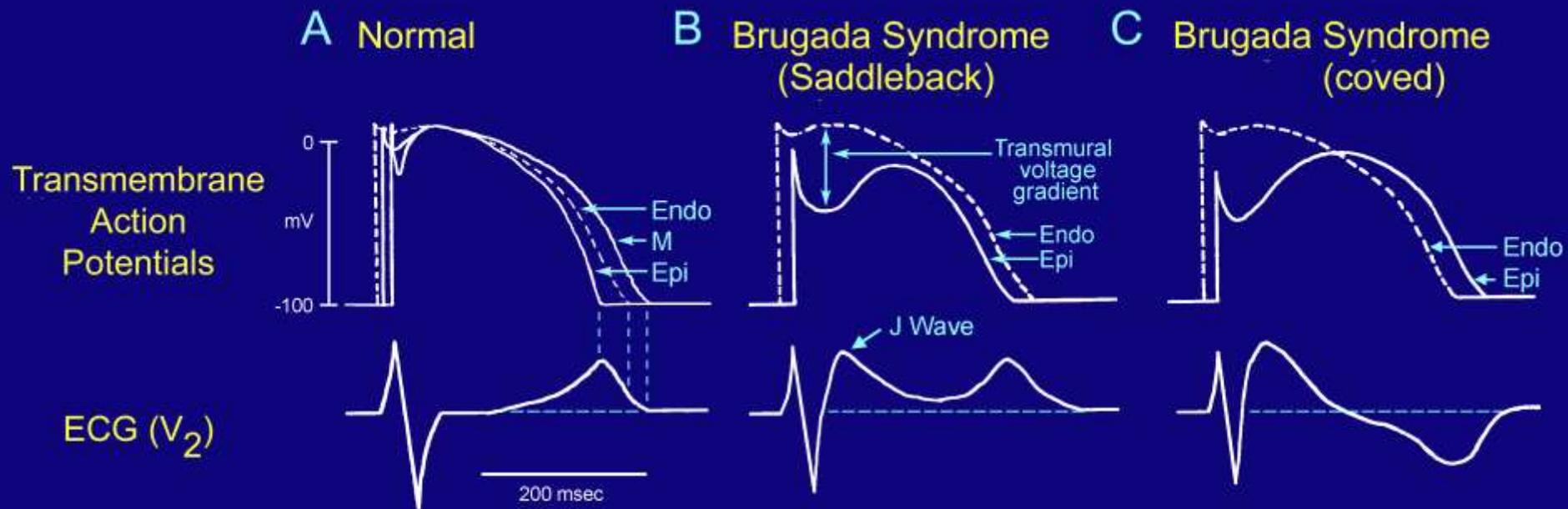
Differences between Brugada and Early Repolarization Syndromes and Possible Underlying Mechanisms

| | BrS | ERS | Possible Mechanism(s) |
|---|--------------------------------|--|---|
| Region most involved | RVOT | Inferior LV wall | Higher levels of I_{to} and/or differences in conduction |
| Leads affected | V1-V3 | II, II aVF V4, V5, V6; I, aVL Both: infero-lateral | |
| Regional difference in prevalence | | | Europe: BrS = ERS Asia: BrS > ERS |
| Incidence of late potential in SAECG | Higher | Lower | |
| Inducibility of VF during an EPS | Higher | Lower | |
| Effect of sodium channel blockers on the surface ECG | Increased J wave manifestation | Reduced J wave Manifestation | Reduction of J wave in the setting of ER is due largely to prolongation of QRS. Accentuation of repolarization defects predominates in BrS, whereas accentuation of depolarization defects predominates in ERS. |

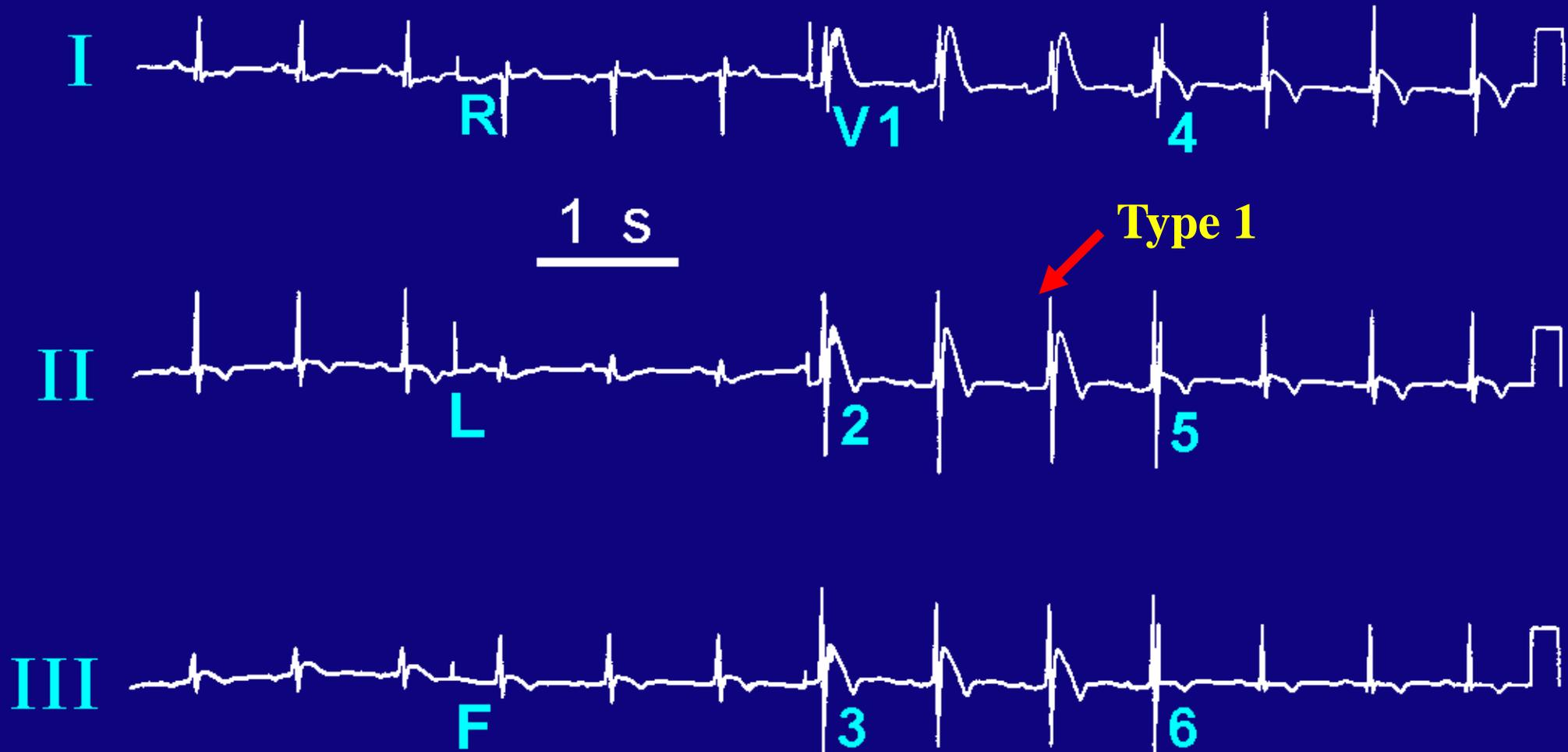
Early Repolarization Patterns



Modified from Antzelevitch et al, JACC, 2011

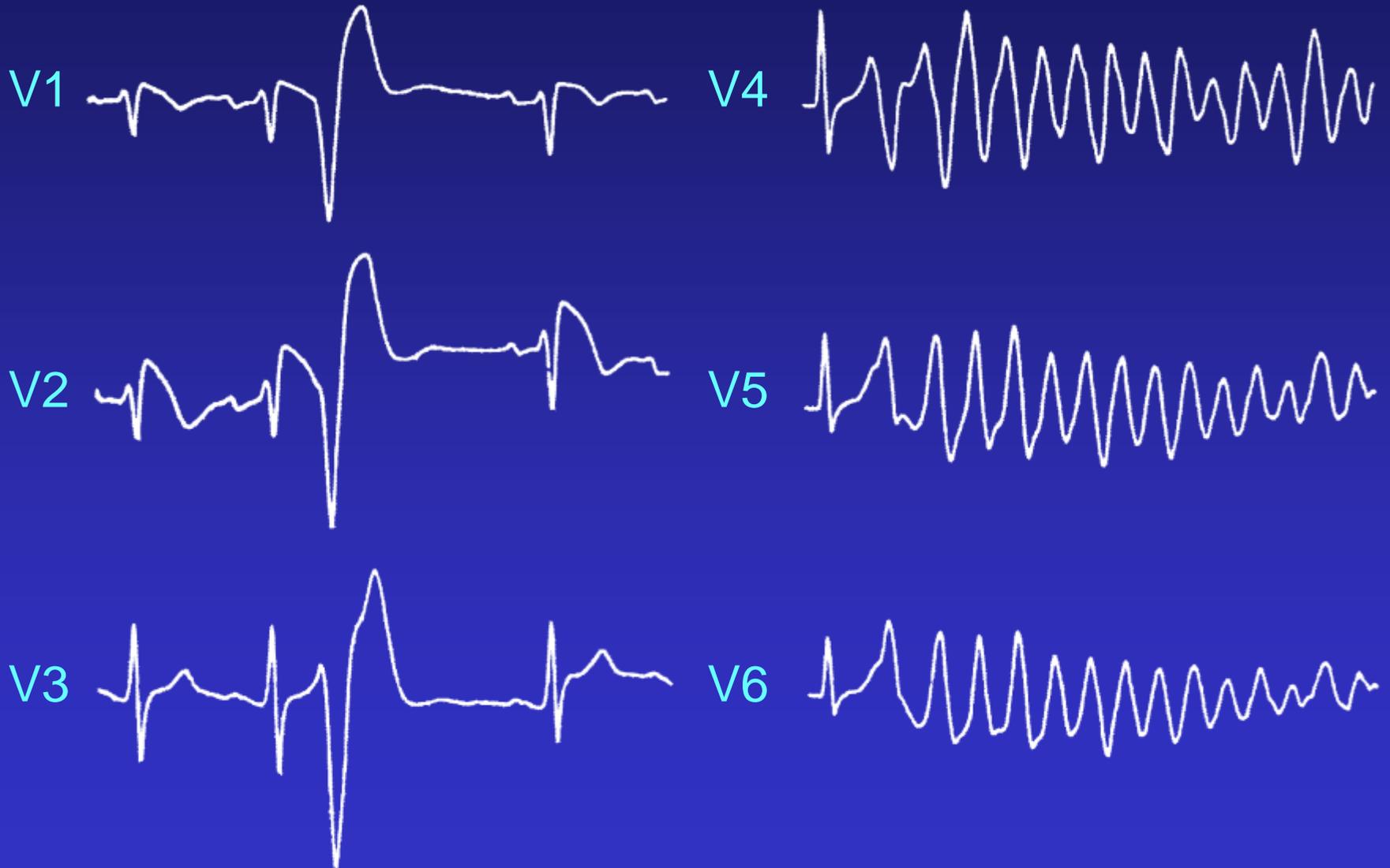


Brugada Syndrome



Brugada and Brugada, JACC 1992;20;1391-96

Ventricular Arrhythmias in Brugada Syndrome



Genetic Basis for Brugada Syndrome - *Causative Genes*

| | Locus | Ion Channel | Gene/Protein | % of Probands |
|--------------|--------------|---|--|---------------|
| BrS1 | 3p21 |  | I_{Na} <i>SCN5A</i> , $Na_v1.5$ | 11-28% |
| BrS2 | 3p24 |  | I_{Na} <i>GPD1L</i> | Rare |
| BrS3 | 12p13.3 |  | I_{Ca} <i>CACNA1C</i> , $Ca_v1.2$ | 6.6% |
| BrS4 | 10p12.33 |  | I_{Ca} <i>CACNB2b</i> , $Ca_v\beta2b$ | 4.8% |
| BrS5 | 19q13.1 |  | I_{Na} <i>SCN1B</i> , $Na_v\beta1$ | 1.1% |
| BrS6 | 11q13-14 |  | I_{to} <i>KCNE3</i> , <i>MiRP2</i> | Rare |
| BrS7 | 11q23.3 |  | I_{Na} <i>SCN3B</i> , $Na_v\beta3$ | Rare |
| BrS8 | 12p11.23 |  | I_{K-ATP} <i>KCNJ8</i> , Kir6.1 | 2% |
| BrS9 | 7q21.11 |  | I_{Ca} <i>CACNA2D1</i> , $Ca_v\alpha2\delta$ | 1.8% |
| BrS10 | 1p13.2 |  | I_{to} <i>KCND3</i> , $K_v4.3$ | Rare |
| BrS11 | 17p13.1 |  | I_{Na} <i>RANGRF</i> , <i>MOG1</i> | Rare |
| BrS12 | 3p21.2-p14.3 |  | I_{Na} <i>SLMAP</i> , Sarcolemma Associated Protein | Rare |
| BrS13 | 12p12.1 |  | I_{K-ATP} <i>ABCC9</i> , <i>SUR2A</i> | Rare |
| BrS14 | 11q23 |  | I_{Na} <i>SCN2B</i> , $Na_v\beta2$ | Rare |
| BrS15 | 12p11 |  | I_{Na} <i>PKP2</i> , Plakophilin-2 | Rare |
| BrS16 | 3q28 |  | I_{Na} <i>FGF12</i> , <i>FHAF1</i> | Rare |
| BrS17 | 3p22.2 |  | I_{Na} <i>SCN10A</i> , $Na_v1.8$ | 5-16.7% |
| BrS18 | 6q |  | I_{Na} <i>HEY2</i> (<i>transcriptional factor</i>) | Rare |
| BrS19 | 1p36.3 | | I_{to} <i>KCNAB2</i> , $K_v\beta2$ | Rare |

Genetic Basis for Brugada Syndrome

New Candidate Genes

| Locus | Ion Channel | Gene/Protein |
|---------|------------------------|--|
| 12p12.1 | ↓ I _{K/Na-Ca} | <i>TRPM4</i> , Transient Receptor Potential Melastatin Protein 4 |
| | ↑ | |
| 7q31.31 | I _{to} | <i>KCND2/Kv4.2</i> |

Modulatory Genes

| | | |
|-----------|-------------------|---------------------------|
| 15q24-q25 | ↓ I _f | <i>HCN4</i> |
| 7q35 | ↑ I _{kr} | <i>KCNH2, HERG</i> |
| Xq22.3 | ↑ I _{to} | <i>KCNE5 (KCNE1-like)</i> |
| 7p12.1 | ↑ I _{to} | <i>SEMA3A, Semaphorin</i> |

Reclassification of pathogenicity of Brugada Syndrome variants classified as deleterious and causative of the inherited syndrome

Recent efforts have led to exhaustive studies of genes and variants previously identified as pathogenic and causative of inherited cardiac arrhythmia syndromes using *American College of Medical Genetics and Genomics and Association for Molecular Pathology guidelines*. This has been performed for BrS led by **Michael Gollob and co-workers**.

The result is that 20 of the 21 genes identified as being causative of the disease have been reclassified as *disputed* with regards to any assertions of disease causality.

Reappraisal of Reported Genes for Sudden Arrhythmic Death: An Evidence-Based Evaluation of Gene Validity for Brugada Syndrome.

Hosseini SM¹, Kim R², Udupa S³, Costain G⁴, Jobling R⁵, Liston E⁵, Jamal SM⁴, Szybowska M⁶, Morel CF⁶, Bowdin S⁵, Garcia J⁷, Care M⁸, Sturm AC⁹, Novelli V¹⁰, Ackerman MJ¹¹, Ware JS¹², Hershberger RE¹³, Wilde AAM¹⁴, Gollob MH¹⁵; NIH-Clinical Genome Resource Consortium.

⊕ Author information

Abstract

Background -Implicit in the genetic evaluation of patients with suspected genetic diseases is the assumption that the genes evaluated are causative for the disease based on robust scientific and statistical evidence. However, in the past 20 years considerable variability has existed in the study design and quality of evidence supporting reported gene-disease associations raising concerns of the validity of many published disease-causing genes. Brugada syndrome (BrS) is an arrhythmia syndrome with a risk of sudden death. More than 20 genes have been reported to cause BrS and are assessed routinely on genetic testing panels in the absence of a systematic, evidence-based evaluation of the evidence supporting the causality of these genes. **Methods** -We evaluated the clinical validity of genes tested by diagnostic laboratories for BrS by assembling three gene curation teams. Using an evidence-based semi-quantitative scoring system of genetic and experimental evidence for gene-disease associations, curation teams independently classified genes as demonstrating Limited, Moderate, Strong or Definitive evidence for disease causation in BrS. The classification of curator teams was reviewed by a Clinical Domain Expert Panel who could modify the classifications based on their independent review and consensus. **Results** -Of 21 genes curated for clinical validity, biocurators classified only 1 gene (SCN5A) as Definitive evidence, while all other genes were classified as Limited evidence. Following comprehensive review by the Clinical Domain Expert Panel, all 20 genes classified as Limited evidence were re-classified as Disputed in regards to any assertions of disease causality for BrS. **Conclusions** -Our results contest the clinical validity of all but one gene clinically tested and reported to be associated with BrS. These findings warrant a systematic, evidence-based evaluation for reported gene-disease associations prior to use in patient care.

Genetic basis and molecular mechanism for idiopathic ventricular fibrillation

Qiuyun Chen, Glenn E. Kirsch, Danmei Zhang, Ramon Brugada, Josep Brugada, Pedro Brugada, Domenico Potenza, Angel Moya, Martin Borggrefe, Gunter Breithardt, Rocio Ortiz-Lopez, Zhiqing Wang, Charles Antzelevitch, Richard E. O'Brien, Eric Schulze-Bahr, Mark T. Keating, Jeffrey A. Towbin & Qing Wang

Ventricular fibrillation causes more than 300,000 sudden deaths each year in the USA alone. In approximately 5–12% of these cases, there are no demonstrable cardiac or non-cardiac causes to account for the episode, which is therefore classified as idiopathic ventricular fibrillation (IVF). A distinct group of IVF patients has been found to present with a characteristic electrocardiographic pattern. Because of the small size of most pedigrees and the high incidence of sudden death, however, molecular genetic studies of IVF have not yet been done. Because IVF causes cardiac rhythm disturbance, we investigated whether malfunction of ion channels could cause the disorder by studying mutations in the cardiac sodium channel gene *SCN5A*. **We have now identified a missense mutation, a splice-donor mutation, and a frameshift mutation in the coding region of *SCN5A* in three IVF families.** We show that sodium channels with the missense mutation recover from inactivation more rapidly than normal and that the frameshift mutation causes the sodium channel to be non-functional. Our results indicate that mutations in cardiac ion-channel genes contribute to the risk of developing IVF.

NATURE 392:294-296, 1998

Genetic Basis for Brugada Syndrome

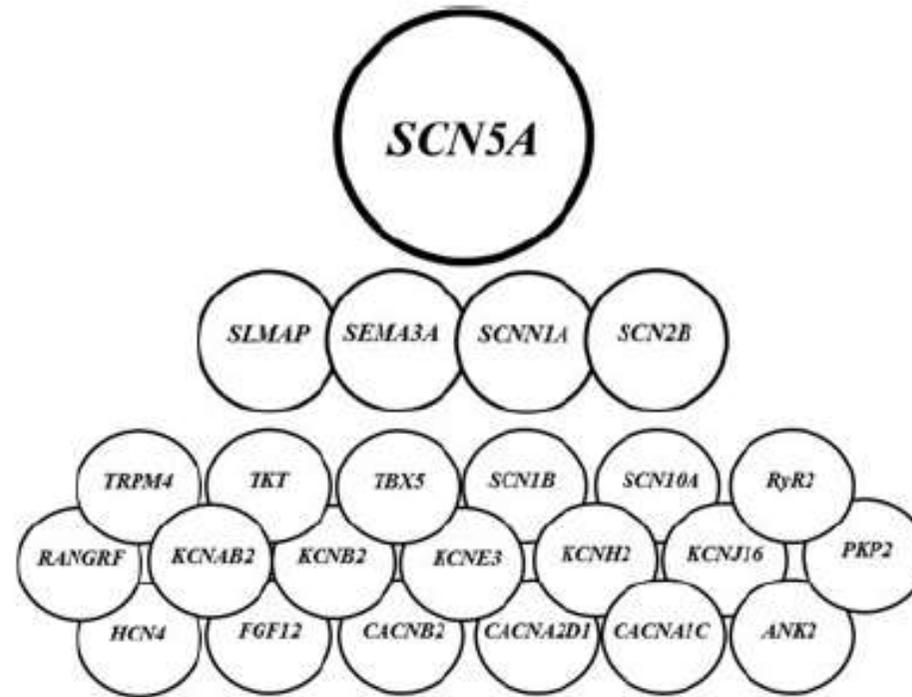
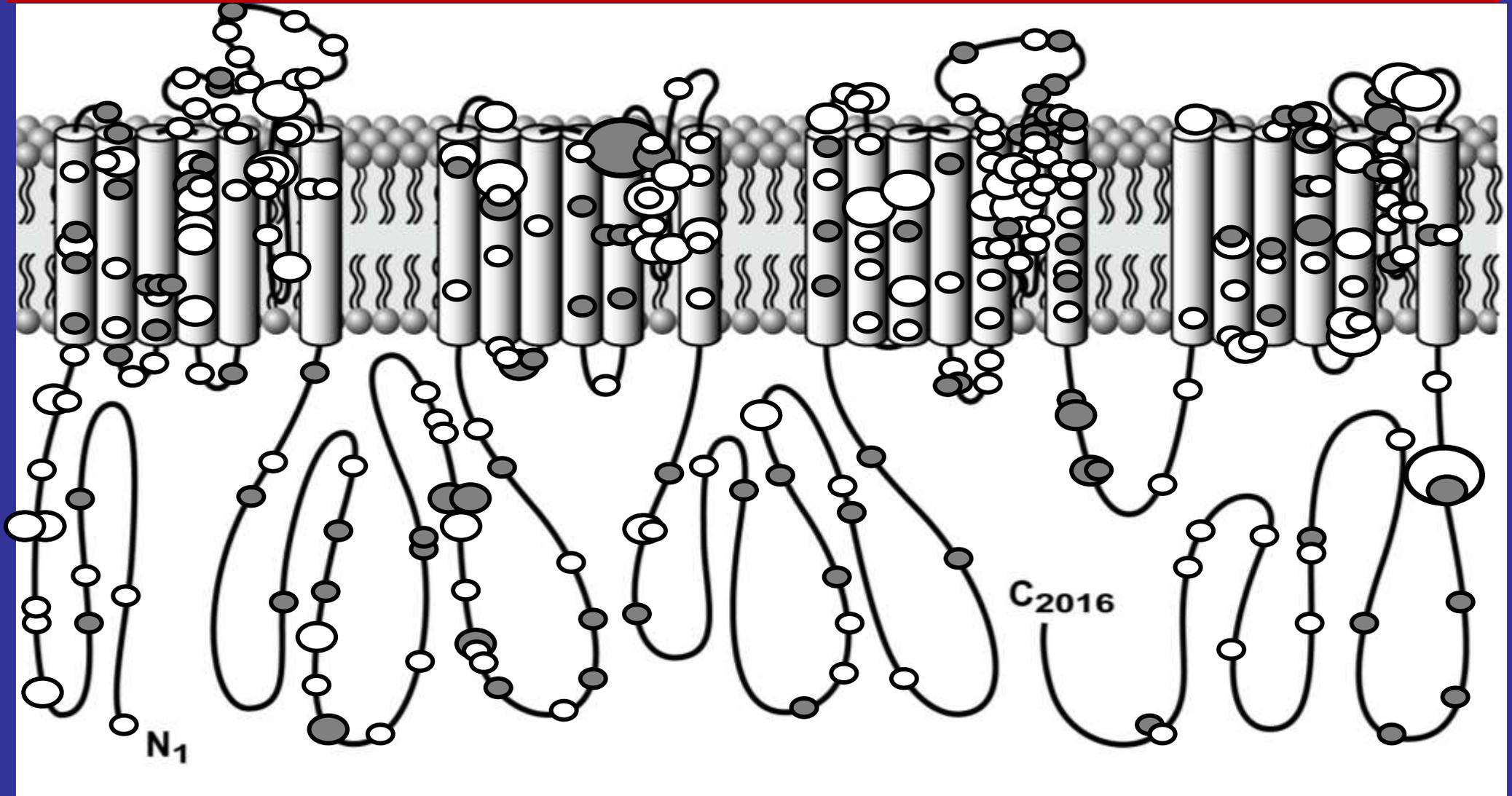
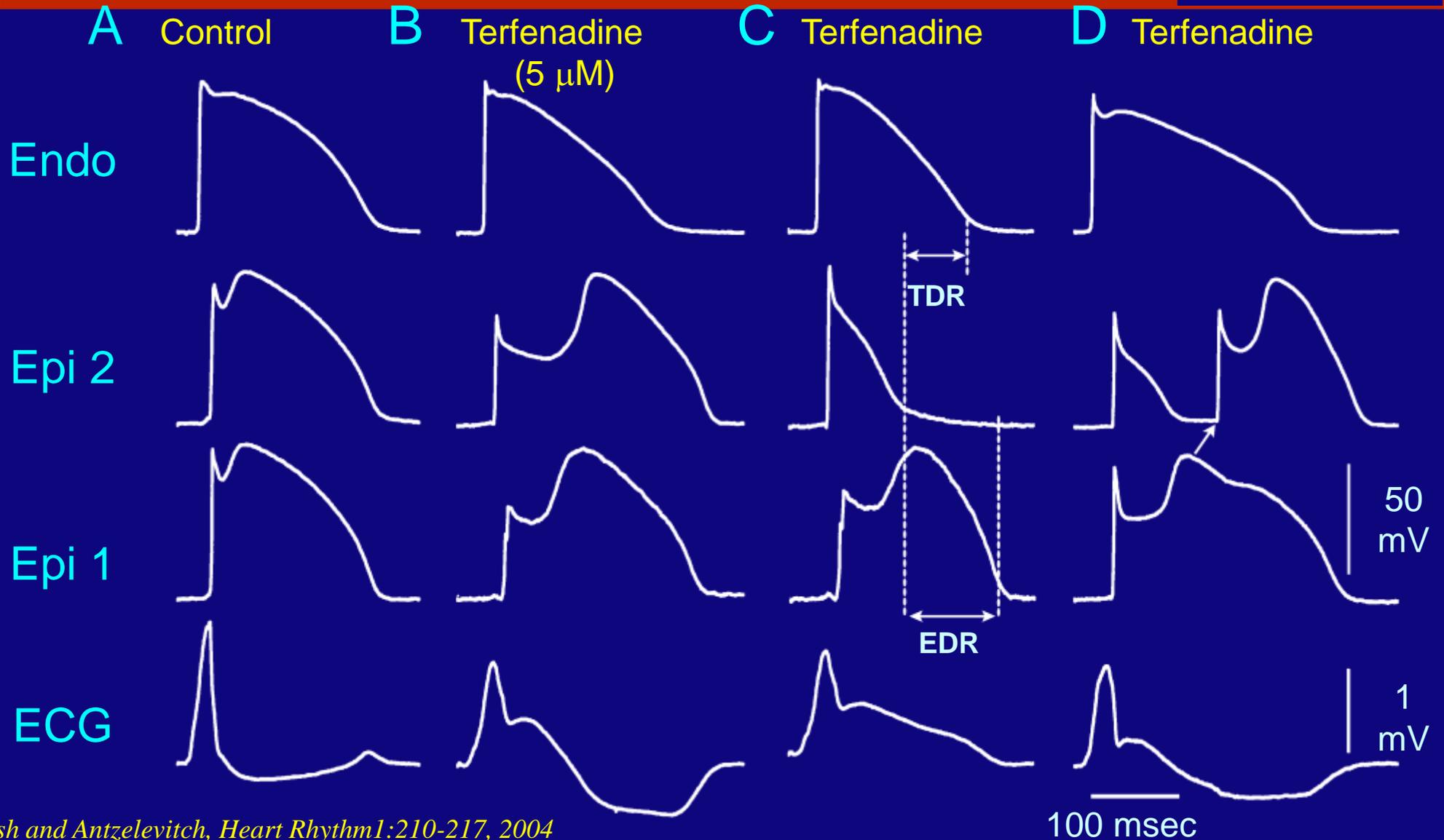
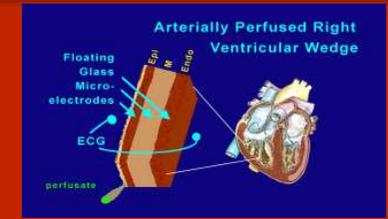


FIGURE 2 Identification of pathogenic and likely pathogenic genes for BrS following ACMG/AMP guidelines. In addition to *SCN5A*, the main gene associated with BrS, 23 other minor genes carry at least one pathogenic or/and likely pathogenic rare variant for BrS. ACMG/AMP: American College of Medical Genetics and Genomics/Association for Molecular Pathology; BrS: Brugada syndrome

Worldwide Brugada Syndrome Consortium
Genetic screening of 211 BrS patients at 9 international centers
293 SCN5A Mutations

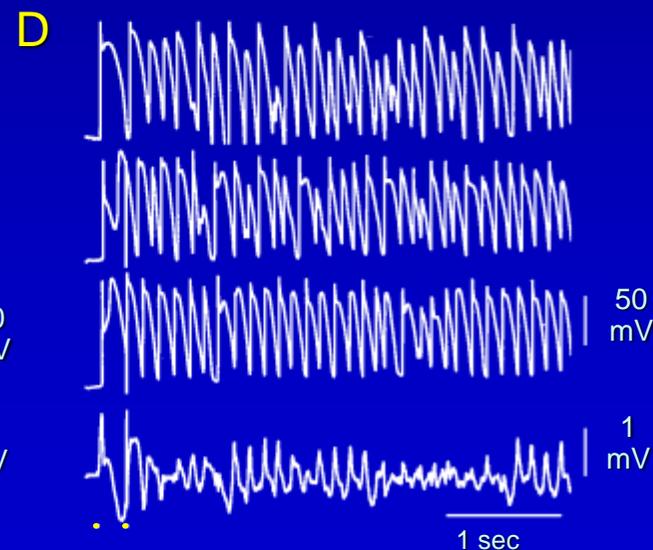
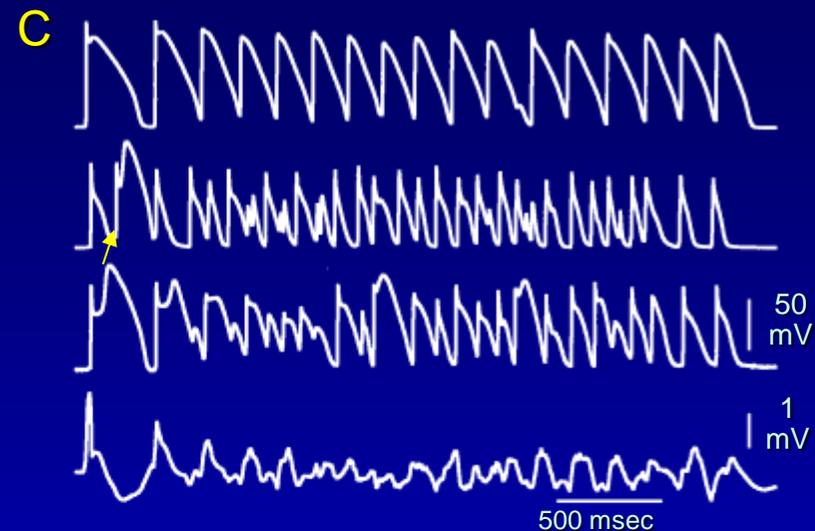
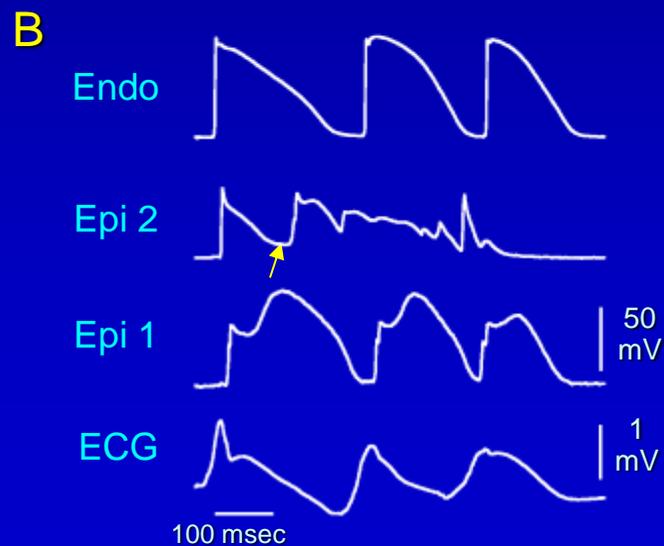
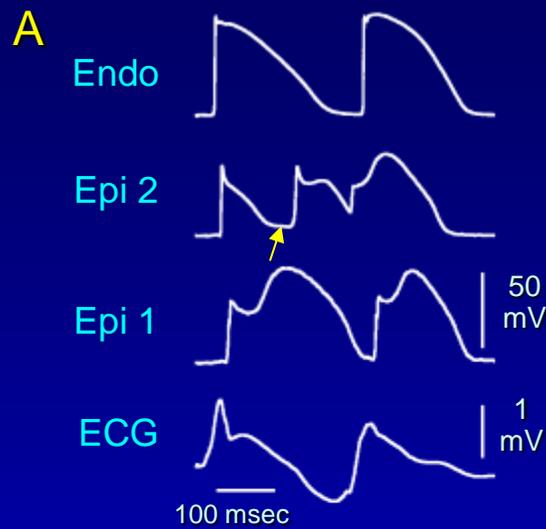
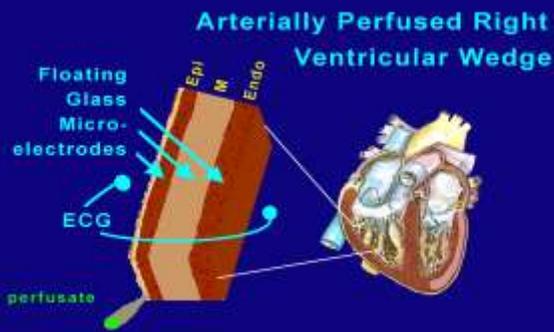


ST Segment Elevation and Phase 2 Reentry Following Combined I_{Na} and I_{Ca} Block

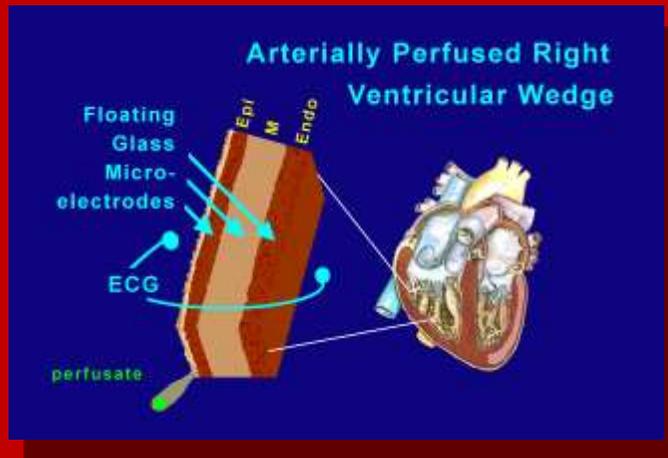


Terfenadine-induced VT/VF

Brugada Syndrome



Early Repolarization Pattern Predisposes to Development of Polymorphic VT/VF via a Brugada Syndrome-like Mechanism

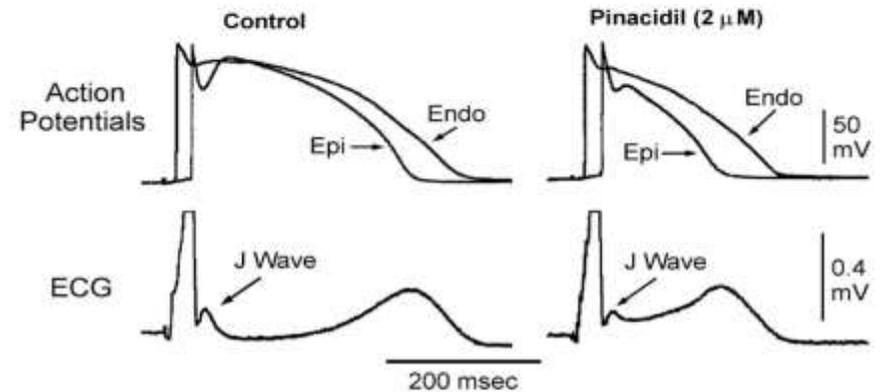


Yan and Antzelevitch, Circulation, 1999
Gussak and Antzelevitch, J Electrocardiol, 2000

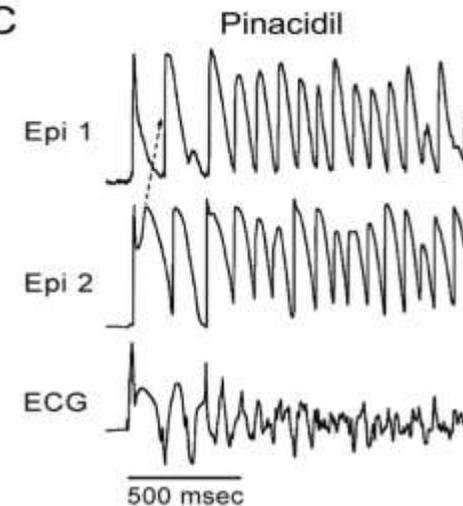
A Early Repolarization Syndrome in a Healthy Young man



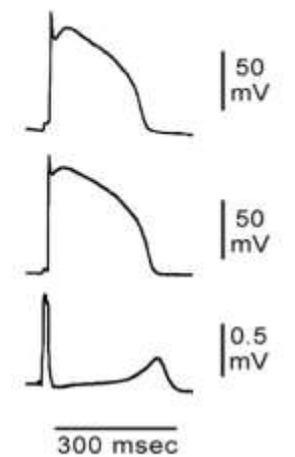
B Canine Ventricular Action Potentials and ECG



C



D + 4-AP



Antzelevitch and Yan, Heart Rhythm 7:549-58, 2010

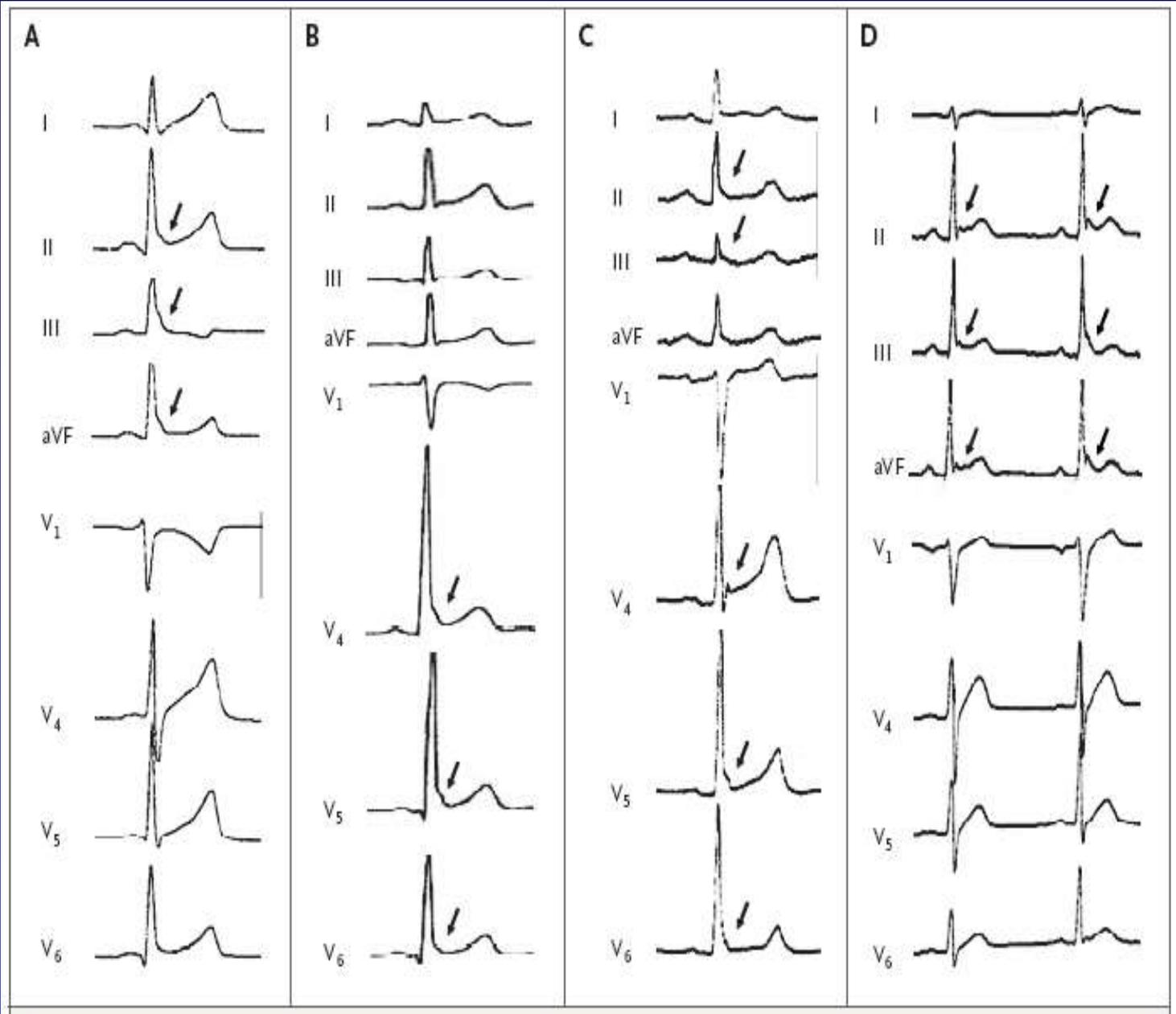
Early Repolarization Pattern and SCD

31% of IVF

vs.

5% of Controls

ER was defined as QRS-ST junction elevation of > 0.1 mV manifested as QRS slurring or notching

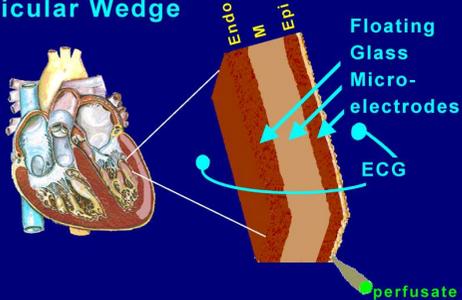


Genetic Basis for Early Repolarization Syndrome

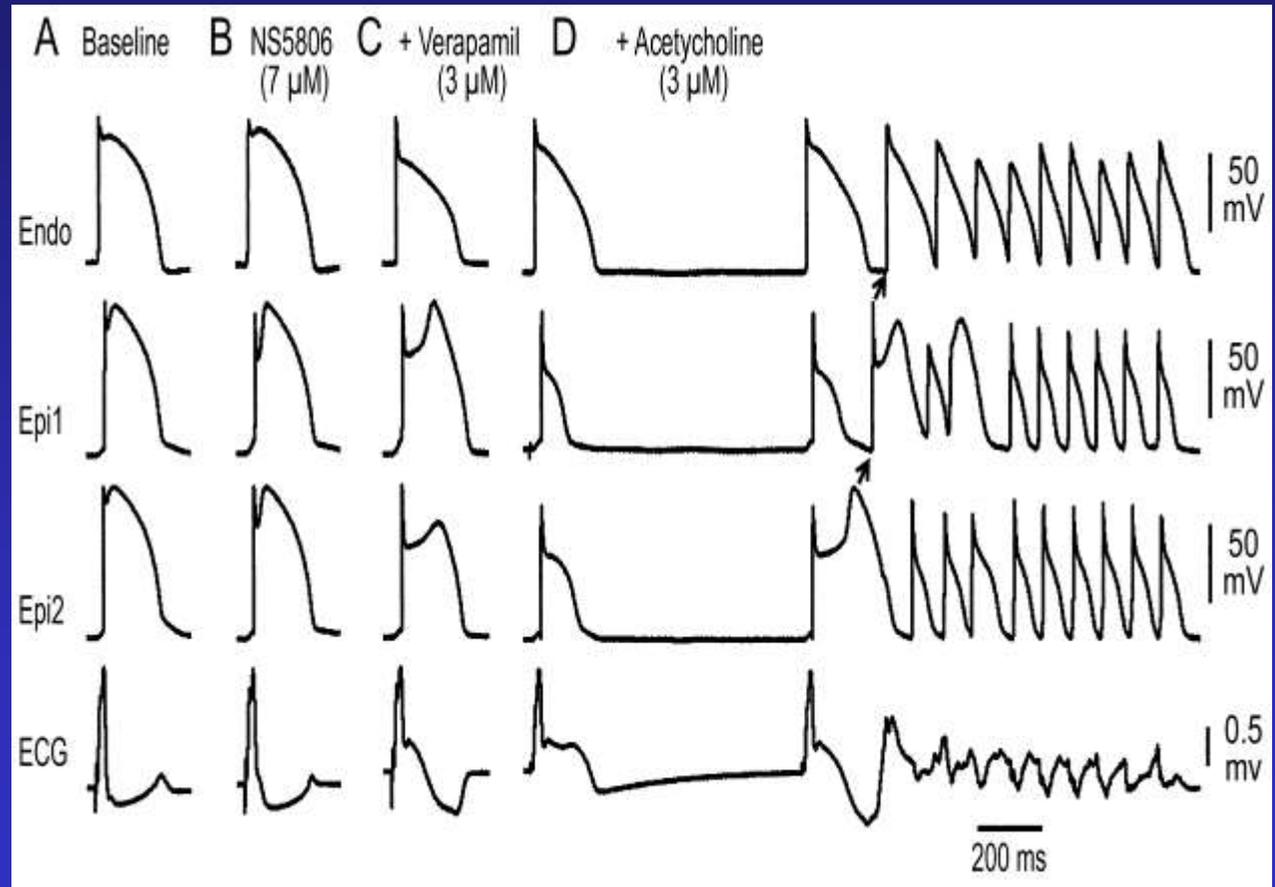
| | Locus | Ion Channel | Gene/Protein | % of Probands |
|------|--------------|----------------------|--|----------------------|
| ERS1 | 12p11.23 | ↑ I _{K-ATP} | <i>KCNJ8, Kir6.1</i> | |
| ERS2 | 12p13.3 | ↓ I _{Ca} | <i>CACNA1C, Ca_v1.2</i> | 4.1% |
| ERS3 | 10p12.33 | ↓ I _{Ca} | <i>CACNB2b, Ca_vβ_{2b}</i> | 8.3 |
| ERS4 | 7q21.11 | ↓ I _{Ca} | <i>CACNA2D1, Ca_vα2d</i> | 4.1% |
| ERS5 | 12p12.1 | ↑ I _{K-ATP} | <i>ABCC9, SUR2A</i> | |
| ERS6 | 3p21 | ↓ I _{Na} | <i>SCN5A, Na_v1.5</i> | |
| ERS7 | 3p22.2 | ↓ I _{Na} | <i>SCN10A, Na_v1.8</i> | |

Cellular Basis for Vagally-mediated potentiation of Early Repolarization Syndrome Phenotype

Arterially Perfused Left Ventricular Wedge

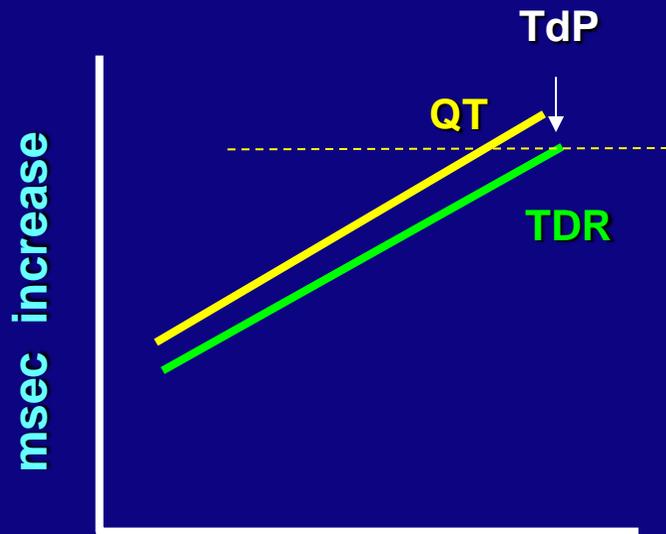


Pharmacologic modeling of I_{CA} loss of function mutations (CACNA1C, CACNB2, CACNA2D1)



Polymorphic VT (PVT)

Long QT Syndrome

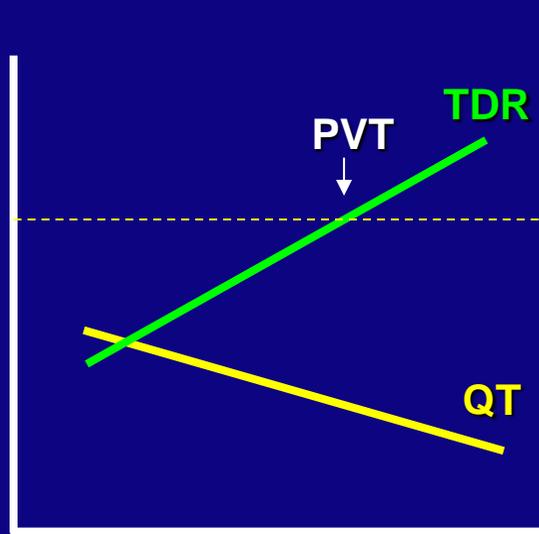


[Drug]
[Disease]

↓ I_{Kr} , I_{Ks} , I_{K1} , I_{K-ACh} ,
↑ I_{Na} , I_{to} , I_{K-ATP}

Dofetilide
Sotalolol, Quinidine

Short QT Syndrome

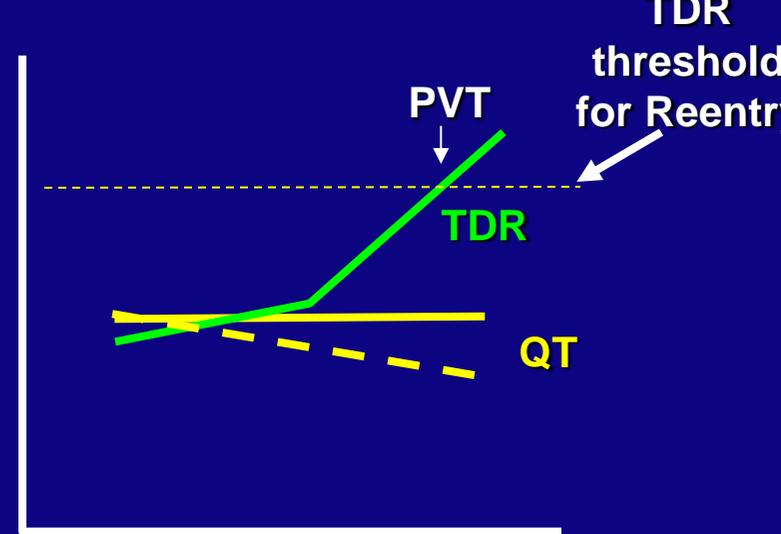


[Drug]
[Disease]

↑ I_{Kr} , I_{Ks} , I_{K1}
↑ I_{K-ATP} , ↓ I_{Ca}

Pinacidil
Digitalis

J Waves Syndromes BrS, ERS



[Drug]
[Disease]

↓ I_{Na} , I_{Ca}
↑ I_{K-ATP} , I_{K-ACh}

Ajmaline, Procainamide

Action Potential Studies

Silvio Litovsky
Anton Lukas
S. Krishnan

Perfused Wedge Studies

Gan-Xin Yan
Serge Sicouri
Wataru Shimizu
Jose Di Diego
A. Burashnikov
Jeffrey Fish
Gi-Byoung Nam
Tetsuro Emori
Fabrice Extramiana
Chinmay Patel
Eyal Nof
Yoshino Minoura
István Koncz
Tamas Szel
Zsolt Gurabai
Bence Patosckai
Namsik Yoon



Voltage Clamp Studies

Helen Diana (Dan Hu)
Hector Barajas-Martinez
Jerome Clatot
Eleonora Savio-Galimberti
Brian Panama
Jonathan Cordeiro

Stem Cell & Molecular Biology

Megan Tabler
Mariana Argenziano
Xavier Michael Jesudoss
Elena Burashnikov
Yuesheng Wu
Mayurika Desai

Molecular Genetics

Jimmy JM Juang
Guido Pollevick
Alejandra Guerchicoff
Ryan Pfeiffer

In Vivo and Modeling Studies

Vladislav V. Nesterenko

Collaborators: Peter Kowey, Andrew Epstein, Sami Viskin, Mel Scheinman, Michel Haissaguerre, Luiz Bellardinelli, Minoru Horie, Yoshifusa Aizawa, Arthur Wilde, Connie Bezzina, Andras Varro, Michael Glickson, Michael Eldar, Liron Miler, Michael Ackerman, Jon Steinberg, Pedro, Josep & Ramon Brugada, Wee Nademanee, Fiorenzo Gaita, Carla Giustetto, Martin Borggreffe, Peter Schwartz, Lia Crotti, Michael Sanguinetti, Mike Ackerman, Christian Wolpert, Rainer Schimpf, Christian Veltmann, Lior Gepstein, Can Hasdemir, Jimmy Juang, Joseph Wu

HRS/EHRA Consensus Statement
(Ackerman et al., Europace 2011)
Expert Consensus Recommendations

| Section # - Disease | Diagnostic | Prognostic | Therapeutic |
|---------------------|------------|------------|-------------|
| • Section I - LQTS | +++ | +++ | ++ |
| • Section III – BrS | + | + | - |
| • Section V – SQTS | +/- | - | - (+) |
| • – ERS | + | ? | - |

- *Relative strength (- = negligible to +++ = strong) of the contribution/impact of the genetic test for diagnosis, prognosis and choice of therapy.*
- **Identification of causative mutations in the proband is important in that it permits the identification of family members who may be at risk and who may require close clinical follow-up.**